

# Peptidyl Arginine Deiminase 4: A Promising Therapeutic Target in Rheumatoid Arthritis and Other Diseases

Murtadha Al-Shaikh Jafar<sup>1</sup>, Hasan Al-Nahab<sup>1</sup>, Mohammad Eid<sup>1</sup>,  
Mohammad Alsamen<sup>1</sup>, Alhassan Almutawah<sup>1</sup>, Mansour Alturki<sup>1</sup>, Nehad Ahmed<sup>2</sup>,  
Mohamed Balaha<sup>2,3</sup>

<sup>1</sup>Department of Pharmaceutical Chemistry, College of Clinical Pharmacy, Imam Abdulrahman Bin Faisal University, Dammam, Saudi Arabia, <sup>2</sup>Department of Clinical Pharmacy, College of Pharmacy, Prince Sattam Bin Abdulaziz University, Al-Kharj, Saudi Arabia, <sup>3</sup>Department of Pharmacology, Faculty of Medicine, Tanta University, Tanta, Egypt

## Abstract

The present review offers comprehensive data on the roles of Peptidyl Arginine Deiminase 4 (PAD4) in various diseases, including rheumatoid arthritis (RA). PAD4, an enzyme responsible for converting arginine to citrulline, has been linked to the development of chronic inflammation. The current study is a narrative review to determine whether PAD4 is a potential therapeutic target for RA and other illnesses. We searched PubMed and Google Scholar for studies published before 2024. The potential therapeutic utility of PAD4 in various diseases such as cancer, miscarriage, multiple sclerosis, retinopathy, and RA is emphasized in the article. This paper explores the advancements in the development of small-molecule inhibitors and monoclonal antibodies targeting PAD4. It stresses the importance of conducting additional research to resolve specificity, safety, and sustained effectiveness concerns. The review emphasizes the need for a comprehensive approach to completely comprehend the varied attributes and potential therapeutic applications of PAD4.

**Key words:** Anti-cyclic citrullinated peptide antibodies, chronic inflammation, peptidyl arginine deiminase 4, rheumatoid arthritis

## INTRODUCTION

Rheumatoid arthritis (RA) is an autoimmune condition where the body attacks its joints, causing inflammation and other symptoms. Genetic and environmental factors like tobacco use can trigger RA, which primarily targets synovial joints, starting with small peripheral joints and often spreading to larger ones.<sup>[1]</sup> Chronic inflammation leads to joint damage, cartilage breakdown, and bone erosion. Early RA lasts less than six months, while established RA persists longer. If untreated, RA worsens over time, increasing the risk of illness and death.<sup>[1]</sup>

Environmental factors, combined with genetic predisposition, can trigger RA in certain individuals. RA patients often have antibodies targeting citrullinated proteins, first identified in 1964 through the antiperinuclear factor. Later, in

1979, anti-keratin antibodies were found, and in the 1990s, these antibodies were shown to specifically target citrullinated filaggrin.<sup>[2,3]</sup> These antibodies, called anti-cyclic citrullinated peptide antibodies (ACPAs), can be identified using cyclic citrullinated peptide (CCP) in enzyme-linked immunosorbent assays (ELISA).<sup>[4]</sup> Citrulline is produced by the enzyme peptidyl arginine deiminase (PAD) at sites of tissue damage and inflammation, such as the lungs in smokers.<sup>[1]</sup>

Patients with a genetic predisposition may produce anti-modified protein antibodies in response to altered proteins.

### Address for correspondence:

Mohamed Balaha, Department of Clinical Pharmacy,  
College of Pharmacy, Prince Sattam Bin Abdulaziz  
University, Al-Kharj 11942, Saudi Arabia.  
E-mail: m.balaha@psau.edu.sa

**Received:** 16-11-2024

**Revised:** 28-02-2025

**Accepted:** 10-03-2025

The enzyme PAD, particularly its isoforms PAD2 and PAD4, is linked to RA by modifying arginine into citrulline. Anti-PAD4 antibodies, specific to RA and associated with ACPA, can be detected in RA patients.<sup>[5]</sup> In gingivitis, the bacterium *Porphyromonas gingivalis* triggers inflammation and leukocyte proliferation, which produces PAD4.<sup>[11]</sup> Peptidyl arginine deiminase 4 (PAD4) is a crucial enzyme for gene expression and the conversion of arginine to citrulline. It governs processes such as cell death, immune defense, and cell differentiation. Dysregulation of PAD4 can lead to various diseases, making it a potential target for the treatment of RA.<sup>[6]</sup>

Peptidyl arginine deiminase 4 (PAD4) is an enzyme that alters proteins by converting arginine to citrulline and aids in the formation of neutrophil extracellular traps (NETs). PAD4 is crucial in the development of cardiovascular diseases, autoimmune disorders, and cancers, making it a potential target for detection and treatment.<sup>[7,8]</sup> Studies have linked PAD expression and citrullination to conditions such as RA, prion disease, psoriasis, Alzheimer's, multiple sclerosis, cancer, and diabetes.<sup>[9]</sup> Jones *et al.* highlight the growing recognition of citrullination's role in RA and other diseases, with PADs, particularly PAD4, identified as promising therapeutic targets.<sup>[10]</sup>

The current study is a narrative review to determine whether peptidyl arginine deiminase 4 (PAD4) is a promising therapeutic target in RA and other disorders. We used Google Scholar and the Trip database to search for studies about PAD 4 and its role in RA and other diseases. Furthermore, for more reference coverage, we went through the references to reviews. The keywords we used were the role of PAD in RA, the role of PAD in other diseases, how PADs work, PAD4 in RA, and PAD4 inhibitors.

## BIOCHEMICAL AND MOLECULAR ASPECTS OF PAD4

Histones are crucial components of chromatin, influencing DNA binding and gene transcription. Posttranslational modifications (PTMs) such as phosphorylation, methylation, acetylation, ubiquitination, and citrullination alter histone function.<sup>[11-13]</sup> Citrullination, facilitated by PAD enzymes, converts peptidyl-arginine to peptidyl-citrulline and is associated with diseases such as RA, Alzheimer's, multiple sclerosis, lupus, Parkinson's, and cancer.<sup>[14,15]</sup> The human PAD family includes five calcium-dependent isozymes (PADs 1-4 and 6), sharing approximately 50% sequence similarity. PADs are distributed across various tissues: PAD1 in the epidermis and uterus, PAD2 in muscle, brain, and secretory glands, PAD3 in hair follicles and keratinocytes, PAD4 in granulocytes and cancers, and PAD6 in oocytes and embryos.<sup>[14]</sup> Although all PADs are cytoplasmic, only PAD4 is known to deiminate histones, with PAD2 potentially playing a role. Recent research indicates that PADs are also

present in granules, mitochondria, and nuclei.<sup>[16,17]</sup> In addition to histones, PADs can target proteins such as fibrinogen, filaggrin, and actin, which are citrullinated in RA.<sup>[18]</sup>

## PAD4 IN RA

In 1998, researchers identified the involvement of the PAD enzyme in RA. Autoantibodies in RA patients recognize citrulline as a crucial autoantigen. The CCP assay detected these as anti-CCPs, or ACPAs, which are vital serological markers for RA diagnosis.<sup>[19]</sup> ACPAs can be detected years before the onset of RA and are associated with preclinical inflammation, severe joint disease, and rapid radiographic progression.<sup>[20]</sup> As ACPAs gained significance in RA research, PAD2, and PAD4 were pinpointed as major drivers of pathogenic citrullination. Studies in mice and humans found these enzymes in hematopoietic cells (granulocytes, monocytes, and macrophages) within the RA synovium, while other PAD isoforms were absent. Deleting or inhibiting PAD2 or PAD4 in mouse models significantly reduced RA severity, underscoring their role in disease progression.<sup>[21-27]</sup>

PAD2 and PAD4 produce citrullinated proteins, such as fibrinogen, vimentin, and  $\alpha$ -enolase, which serve as autoantigens in RA.<sup>[28,29]</sup> However, the precise role of each PAD enzyme in contributing to the RA citrullinome remains uncertain. While PAD2 and PAD4 preferentially citrullinate different but partially overlapping substrates, neither enzyme alone can generate all the citrullinated antigens observed in RA.<sup>[30-32]</sup> These studies reveal distinct citrullination patterns, which may lead to autoantigens with varying immunogenicity. ACPAs in the blood of RA patients bind to citrullinated fibrinogen from both PAD2 and PAD4 at low concentrations, but at high concentrations, they favor PAD4-citrullinated fibrinogen.

PAD enzymes are pivotal in the aberrant citrullination observed in RA, leading to autoimmunity against citrullinated antigens. This dual role makes PAD enzymes both contributors to and targets of the autoimmune response.<sup>[5]</sup> Disrupted citrullination can cause the immune system to attack PAD enzymes, potentially initiating a cycle that leads to key RA symptoms: generation of citrullinated autoantigens, release of pro-inflammatory cytokines, and joint damage. Some RA patients have autoantibodies targeting PAD4, PAD2, or both, which correlate with different clinical outcomes, indicating that anti-PAD antibodies could serve as predictive biomarkers.<sup>[5]</sup> Incorporating these antibodies alongside ACPAs, RF, and other markers could aid in better categorizing RA patients. Anti-PAD4 antibodies can either intensify or mitigate joint damage in RA, depending on their specific epitopes.<sup>[28,33]</sup> Ferucci *et al.* found that anti-PAD4 antibodies are more prevalent in RA patients than in first-degree relatives without RA, suggesting they emerge earlier than anti-CCP antibodies in RA development.<sup>[34]</sup> Studies also associate high levels of PAD, citrullination, and related diseases, including RA, prion

disease, psoriasis, Alzheimer's, multiple sclerosis, cancer, and others.<sup>[9]</sup>

## ROLE OF PAD4 IN DIFFERENT DISEASES

Studies have explored PAD's role in glioblastoma multiforme (GBM), where PAD pathways influence the formation and release of extracellular vesicles (EVs) in GBM and other cancers.<sup>[35]</sup> These EVs facilitate tumor growth, angiogenesis, and invasion. A PAD3 inhibitor reduced GBM cell invasion and altered EVs in LN229 cells, though PAD4 and PAD2 inhibitors were more effective in LN18 cells.<sup>[35]</sup>

Elevated PAD4 expression is associated with chemotherapy resistance in hepatocellular carcinoma (HCC) patients undergoing transcatheter arterial chemoembolization post-surgery. The overproduction of PAD4 in HCC cells induces autophagy, a protective mechanism against cytotoxicity, leading to chemotherapy resistance.<sup>[36,37]</sup> Exosomes and microvesicles are crucial in intercellular communication, transporting molecules such as cytokines, growth factors, and miRNAs that influence processes such as differentiation, migration, and angiogenesis.<sup>[36-39]</sup> In addition, PAD2 and PAD4 have been shown to translocate to the nucleus in response to TNF overexpression.<sup>[40-42]</sup>

Renal insufficiency affects about 50% of cancer patients at diagnosis, leading to reduced effectiveness of anticancer treatments due to lowered medication dosages. In mice models, kidney function showed decreased creatinine clearance and increased urine protein levels. Electron microscopy revealed reversible changes, such as increased mesangial cells, but no permanent damage such as fibrosis or necrosis. Treatment with DNase I or PAD4 inhibitors restored renal function by eliminating NETs. PAD4 inhibitors may also help reduce tumor-induced systemic inflammation and improve blood flow in peripheral arteries.<sup>[43]</sup> The PAD4 gene is linked to epigenetic and phenotypic changes in several cancers.<sup>[44-46]</sup> Increased PAD4 expression enhances the malignant properties of NPC cells. GSK484 reduces citH3 protein levels, a marker of PAD4 activity, and inhibits NPC progression by suppressing PAD4's citrullination.<sup>[47]</sup> Since GSK484 is a reversible inhibitor, extended exposure may not entirely inactivate PAD4. Nevertheless, GSK484 notably inhibited tumor growth *in vivo*, suggesting PAD4 as a potential biomarker for NPC treatment. F-amidine, with its ability to irreversibly inhibit PAD4, may offer benefits over GSK484.<sup>[47]</sup> Furthermore, PAD4 is highly expressed in stomach cancer, driving cell proliferation, and its inhibitors could help prevent cancer metastasis.<sup>[46]</sup>

Inflammation is a key factor in placentation problems, including miscarriages, fetal growth restriction, and preeclampsia.<sup>[48]</sup> In pregnant mice, overproduction of soluble Flt-1 (sFlt-1) leads to issues ranging from spontaneous fetal loss to preeclampsia, with the severity dependent on the

dosage.<sup>[49]</sup> Alterations in sFlt-1 and other angiogenic proteins, such as placental growth factor, have been associated with various reproductive conditions, including miscarriages, fetal growth limitation, and preeclampsia.<sup>[50-52]</sup> Sarcoma-like tyrosine kinase 1 (sFlt-1) induces significant neutrophil accumulation and NET formation in mouse placentas. In a mouse model with sFlt-1 overexpression, the absence of PAD4 reduces both placental and systemic inflammation, protecting against pregnancy loss caused by sFlt-1.<sup>[53]</sup> The pregnancy loss linked to sFlt-1 is associated with PAD4-triggered NETosis. Thus, inhibiting PAD4 may offer a novel strategy to prevent inflammatory miscarriages during early placentation defects, potentially preserving pregnancies.

Retinal stress and disease induce excessive production of glial fibrillary acidic protein (GFAP), which undergoes citrullination in the damaged retina.<sup>[54,55]</sup> Inhibiting PAD4-mediated citrullination results in a significant decrease in GFAP expression, indicating that targeting GFAP could be a promising treatment strategy for retinopathy.<sup>[56]</sup> PAD4 is crucial in inflammation during ischemic acute kidney injury (AKI), promoting renal tubular inflammation, neutrophil infiltration, and NFκB activation.<sup>[57]</sup> In mice, PAD4 activity increases following renal ischemia-reperfusion (I/R) injury. After 30 minutes of ischemia, untreated mice exhibited severe AKI with elevated plasma creatinine levels. Pretreatment of mice with PAD4 inhibitors (2-chloroamidine or streptonigrin) led to a significant reduction in I/R injury.<sup>[58]</sup> These findings emphasize PAD4's role in enhancing inflammatory responses and neutrophil infiltration during renal I/R injury.<sup>[58]</sup> Liver I/R injury initiates the release of damage-associated molecular patterns, triggering an immune response. Treatment with PAD4 inhibitors or DNase I significantly protects hepatocytes and reduces inflammation by inhibiting NET formation, underscoring their role in liver I/R injury. PAD4 inhibitors and DNase I reduce liver damage mediated by High Mobility Group Box 1 and histones.<sup>[59]</sup> The PAD4 inhibitor YW4-03 notably decreased liver I/R injury, suggesting its potential as a therapeutic agent. Targeting NETs with PAD4 inhibitors or DNase I may improve outcomes in liver surgery patients at risk of ischemia/reperfusion injury.<sup>[59]</sup>

In systemic lupus erythematosus (SLE), patients have an impaired ability to break down NETs, which can result in increased exposure of nuclear antigens to the immune system. Suppressing PAD activity significantly alleviates vascular, renal, and skin disease in lupus-prone mice by reducing NET formation.<sup>[60,61]</sup> However, PAD4 deletion in glomerulonephritis models did not improve nephritis or immune activation, and PAD inhibition had no effect on end-organ damage. Treatment of mice in New Zealand with Cl-amidine reduced NET formation, altered autoantibody profiles, decreased complement levels, and lowered IgG deposition in the glomeruli, suggesting potential therapeutic benefits in SLE.<sup>[62,63]</sup>

Cl-amidine not only reduced NET formation but also enhanced bone marrow endothelial progenitor cell differentiation,

improved endothelial-dependent blood vessel relaxation, and delayed arterial thrombosis after photochemical exposure.<sup>[63]</sup> In SLE, dysfunctional NET degradation leads to an accumulation of NETs, which can trigger Type I IFN responses, activate inflammasomes, and cause endothelial damage, contributing to abnormal immune responses.<sup>[64-66]</sup> These findings suggest that inhibiting PAD activity may help modulate the development and severity of lupus and its symptoms.

Elevated amounts of ACPAs have been seen in the bloodstream of a xenografted animal model of ulcerative colitis.<sup>[67]</sup> Therefore, it is hypothesized that the development of ulcerative colitis may be attributed to an excessive level of citrullination. In Alzheimer's disease (AD), PADs and citrullinated proteins are more prevalent in brain regions affected by neurodegeneration. Neuronal death releases citrullinated proteins into the interstitial space, which can enter the bloodstream and lymphatic system, triggering an immune response. This results in persistent exposure of the central nervous system to the immune system.<sup>[68]</sup>

PAD4, citrullinated peptides, and NETs have been found in atherosclerotic plaques, suggesting that immune complexes at these sites may drive inflammation and disease progression.<sup>[69,70]</sup> A study of 134 female RA patients showed a strong link between ACPA levels and subclinical atherosclerosis. Recent research further confirmed NETs' role in atherosclerosis, showing that cholesterol crystals trigger NETosis, IL-1b production, and TH17 activation, which recruit immune cells.<sup>[70]</sup> In mouse models, PAD inhibition reduced NET formation, lesion size, and thrombosis risk while also decreasing immune cell recruitment and interferon- $\alpha$  production in arteries.<sup>[71]</sup> Inhibiting PAD4 could thus reduce NET formation and immune activation, offering a potential therapeutic approach.

Protein citrullination was once thought to be a bacterial strategy to evade immune detection by deactivating NETs.<sup>[72]</sup> However, experiments showed that inhibiting PAD4 had no effect on amoebic NETosis, while calcium chelation inhibited the process, as calcium is crucial for neutrophil adhesion during NETosis.<sup>[73]</sup> The PAD4 inhibitor GSK484 also failed to suppress NET release triggered by amoebas, suggesting that while citrullination plays a role in NETosis, it may not be involved in amoebic NETosis, which likely occurs via a different mechanism.<sup>[74]</sup> In contrast to *Staphylococcus*, PAD4 inhibition reduced phagocytosis and oxidase activation.<sup>[75]</sup> PAD2 levels are markedly higher in the L $\beta$ T2 gonadotrope cell line compared to other PAD isoforms, with peak expression occurring during estrus. This indicates that PADs may be involved in regulating gonadotropin synthesis in females, potentially offering a new approach to controlling fertility through PAD-catalyzed citrullination in gonadotropes. The functional overlap between PAD2 and PAD4 may explain why PAD2 or PAD4 knockout mice have limited utility in reproductive studies, as they seem to compensate for each other's roles.<sup>[76,77]</sup>

NETs facilitate thrombosis by serving as a scaffold for fibrin deposition and enhancing platelet adhesion, aggregation, and activation. Mice deficient in PAD4 are protected from DVT in a stenosis model, with no extracellular citrullinated histones detected despite neutrophil mobilization. Infusion of wild-type neutrophils restored thrombosis, highlighting the crucial role of PAD4 in neutrophils for thrombosis. Advanced imaging has replicated DVT dynamics in mice, showing that high interstitial hemodynamic forces trigger NET release in sterile occlusive thrombosis, though PAD4 inhibition did not prevent NETosis.<sup>[78-80]</sup> PAD4 in the bloodstream impairs the clearance of VWF-platelet strings and accelerates platelet plug formation after vessel damage, partly by reducing ADAMTS13 activity.<sup>[81,82]</sup> The enzyme r-huPAD4 converts specific arginine residues on ADAMTS13 into citrulline, significantly decreasing its activity. Plasma samples from patients with sepsis or elderly individuals with comorbidities (e.g., diabetes, hypertension) show increased citrullination of ADAMTS13 compared to healthy controls.<sup>[83]</sup>

NET levels are elevated in nonhealing diabetic foot ulcers. In mice, diabetes increases skin PAD4 activity, leading to histone citrullination. Inhibiting PAD4 with Cl-amidine reduced neutrophil-derived NETs and improved wound healing. Cl-amidine may, therefore, be a promising topical treatment for diabetic individuals.<sup>[84]</sup> YW3-56, a PAD2/PAD4 inhibitor, reduces lipopolysaccharide (LPS)-induced pulmonary dysfunction by decreasing NET formation and CitH3 generation. This compound helps preserve endothelial cell integrity, reduces pulmonary vascular dysfunction, and alleviates acute lung injury (ALI), improving survival in a mouse model of endotoxemia.<sup>[85]</sup> In another experiment, TDFA, a PAD4-specific inhibitor, reduced lung edema, pulmonary damage, and mortality after LPS exposure. TDFA also suppressed inflammation by lowering pro-inflammatory cytokines and oxidative stress, suggesting a potential novel treatment for LPS-induced ALI.<sup>[86]</sup>

Mice lacking PAD4 and infected with bacteremia showed similar survival rates to normal mice, with no worse outcomes. Antibiotic treatment in PAD4-deficient mice slightly increased the death rate, but bacterial presence in the bloodstream remained unchanged, and there was partial protection against LPS-induced shock.<sup>[87]</sup> These findings suggest that PAD4/NETs-mediated toxic inflammation is driven by bacterial endotoxins, and inhibiting PAD4 to prevent NET formation in inflammatory disorders is unlikely to increase infection susceptibility.<sup>[87]</sup> A study of 127 patients found that high PAD4 levels, but not specific PAD4 polymorphisms (PAD4\_89, PAD4\_94, PAD4\_104), were linked to increased mortality in septic shock patients in the ICU.<sup>[88]</sup>

PAD4 influences key transcription factors like NF $\kappa$ B and interacts with the apoptotic regulator p53, playing a role in inflammatory gene production.<sup>[89]</sup> It also regulates CXCL chemokine production in human macrophages, especially

when stimulated by TLR ligands or *Plasmodium falciparum*. CXCL1 may serve as a biomarker for severe malaria. Beyond neutrophil regulation, PAD4 controls chemokine synthesis in macrophages, with its inhibition delaying liver cell movement. While blocking PAD4 doesn't significantly affect individual chemokines, its overall impact on multiple chemokines may explain this delay. PAD4's role in macrophage death could further explain the observed delays in CXCL1 and CXCL2 production when PAD4 is inhibited.<sup>[90,91]</sup>

Patients with acute pulmonary embolism (APE) display elevated levels of NETs and lysophosphatidic acid (LPA). Autotaxin, the enzyme responsible for producing LPA, is present in intrapulmonary thrombi and surrounds NETs. LPA rapidly induces NET release through a PAD4-dependent pathway, and these LPA-induced NETs contribute to a thrombus that is resistant to tissue plasminogen activator (tPA). This suggests that PAD4 inhibitors may be necessary to overcome tPA resistance in APE.<sup>[92]</sup> PAD4 is crucial for NET formation and the release of DNA in neutrophils. In individuals with cystic fibrosis (CF), higher levels of anti-PAD4 antibodies are linked to lung infections caused by *Pseudomonas aeruginosa*. Neutrophils are the primary source of PAD4 in CF, as they are abundant in the airways, express high levels of PAD4, and play a key role in NET release.<sup>[93,94]</sup>

PAD2 and PAD4 enzymes are present in the salivary glands of Balb/c mice, but their ability to citrullinate Ro and La ribonucleoproteins remains uncertain. In Sjögren's syndrome, PTMs in the salivary glands may rely on PAD activity. These modifications suggest that external triggers, such as bacteria or chemical compounds, can activate the PAD pathway, transforming normal proteins into potential autoantigens. This process may stimulate autoimmune class II MHC molecules and autoreactive cell clones, initiating an autoimmune response akin to Sjögren's syndrome.<sup>[95]</sup>

*P. gingivalis*, linked to severe periodontitis, secretes an enzyme that converts peptidyl arginine into citrulline residues. To survive the influx of neutrophils in inflamed gum tissue, *P. gingivalis* uses Porphyromonas Peptidylarginine Deaminase (PPAD) to prevent neutrophil death. PPAD also modifies proteins, including certain integrins, to facilitate their uptake by phagocytes. If bacterial PPAD shares homology with human PADs, inhibitors of human PADs could be repurposed as antibiotics. However, if *P. gingivalis* has a distinct active site, adapting existing PAD inhibitors could offer a novel approach to targeting this pathogen.<sup>[96]</sup>

## THERAPEUTIC TARGETING OF PAD4 IN RA

Cells of myeloid lineage, including neutrophils, eosinophils, and monocytes/macrophages, express PAD4, which is believed to regulate gene expression, cell division, programmed cell death, NETs, and tumor development. PAD4 plays a crucial role in cardiovascular diseases, autoimmune disorders,

and cancers, making it a promising target for diagnosis and treatment.<sup>[97]</sup> Small-molecule inhibitors of PAD, such as BB-Cl-amidine and the PAD4-specific GSK199, show therapeutic potential and aid in citrullination research. Using an ELISA assay, Martín Monreal *et al.* discovered that AFM-30a effectively inhibited PAD activity in intact PMNs, lysed PBMCs, and synovial fluid from RA patients, while GSK199 had minimal effect.<sup>[98]</sup> In addition, AFM-30a effectively inhibits PAD2 in PBMCs, PMNs, and synovial fluid, as demonstrated by Martín Monreal *et al.* When combined with GSK199, AFM-30a reduces PAD activity in PBMCs without the cytotoxicity observed with BB-Cl-amidine. These findings suggest that AFM-30a and GSK199 may have fewer side effects, making them potentially more suitable for therapeutic use.<sup>[98]</sup>

A preliminary study indicates that anti-PAD4 monoclonal antibodies may enhance the production of pro-inflammatory cytokines in monocytes, potentially transforming them into bone-resorbing cells, a process associated with RA.<sup>[99]</sup> The precise mechanism involving autoantibodies remains unclear. However, the presence of PAD4 on monocyte surfaces and the binding of ACPAs to immature DC-derived osteoclasts suggest that these antibodies could initiate pathogenic effects.<sup>[99,100]</sup> This interaction may activate NK cells, inducing antibody-dependent cytotoxicity through perforin, granzyme, or complement activation, leading to sustained histone release and the production of citrullinated RA autoantigens by neutrophils.<sup>[101]</sup>

Comprehending immune responses to citrullinated proteins and PAD enzymes could pave the way for novel treatments that decouple inflammatory and degenerative effects from PAD enzyme function. Researchers have suggested using chimeric autoantibody receptor T-cell vaccines to target specific antibodies in autoimmune disorders.<sup>[102]</sup> In a safety trial, Benham *et al.* discovered that Rheumavax, a personalized dendritic cell vaccine aimed at inducing tolerance to citrullinated peptides, exhibited immunomodulatory and anti-inflammatory effects in RA patients.<sup>[103]</sup> With a deeper understanding of the specific immunogenic epitopes involved in RA, targeted immunotherapies could be developed to suppress autoreactive T and B cells while preserving PAD enzyme function.<sup>[5]</sup> Antibodies targeting PAD4 and PAD2 have been identified in certain groups of RA patients, and these autoantibodies could serve as predictive biomarkers. Incorporating anti-PAD antibodies alongside ACPAs, RF, and other autoantibodies could aid in better categorizing RA patients. However, further research is needed to fully understand the regulation of PAD4 activity.<sup>[5]</sup> Liu *et al.* emphasized the importance of developing PAD4 inhibitors, as PAD4 is implicated in multiple disorders. Despite advances in creating PAD4 inhibitors targeting specific isotypes, none are currently available for clinical use.<sup>[104]</sup> Further research into PAD4's structure, function, and regulation is crucial for designing effective inhibitors to treat autoimmune and inflammatory diseases.

## POTENTIAL SIDE EFFECTS AND SAFETY CONCERNS

PAD4 inhibitors offer potential for treating various conditions, but like any medication, they may have side effects. Still in the experimental phase, their safety and effectiveness have not been fully established.<sup>[36]</sup> Research has shown that mice without PAD are more prone to bacterial infections, indicating that inhibiting PAD activity and NET formation could increase infection risk.<sup>[36]</sup> Thanabalasuriar *et al.* found that PAD4-deficient mice exhibited extensive inflammation and bacterial keratitis, an eye infection.<sup>[105]</sup>

Lu *et al.* used the PAD4 inhibitor YW3-56 with an Au nanodrug delivery system to target tumors and induce cell apoptosis through chemical-photothermal therapy, significantly inhibiting tumor growth and lung metastasis in mice with good biosafety.<sup>[106]</sup> Gajendran *et al.* also found that the PAD4 inhibitor JBI-589 did not negatively impact body or spleen weight in mice.<sup>[107]</sup> However, no PAD4 inhibitor has been approved for human use, emphasizing the need for further development of inhibitors with optimal ADME properties, safety, and efficacy.<sup>[107]</sup> Yu *et al.* created both reversible (e.g., GSK199 and GSK484) and irreversible (e.g., F- and Cl-amidine) PAD4 inhibitors effective against inflammation and cancer, which are in preclinical stages.<sup>[80]</sup> Future research should aim to discover safer and more effective drugs for clinical use, with a thorough understanding of PAD4 to avoid side effects like increased infection risk. Yang *et al.* showed that GSK199 was effective in preventing murine arthritis.<sup>[7,8]</sup> To minimize infection risks, future PAD4 inhibitors should be designed to act extracellularly and avoid entering cells.

## CONCLUSION

In conclusion, this review highlights the significance of PAD4 as a promising therapeutic target for RA and other related conditions. PAD4's role in citrullination and the formation of autoantibodies underscores its potential for targeted intervention in RA and associated diseases. While targeting PAD4 offers significant therapeutic promise, future research and clinical trials must address challenges related to specificity, safety, and long-term efficacy. A comprehensive understanding of PAD4's diverse functions, beyond its enzymatic activity, is essential to fully explore its therapeutic potential.

## ACKNOWLEDGMENTS

This study is supported via funding from Prince Sattam bin Abdulaziz University project number (PSAU/2024/R/1445).

## REFERENCES

1. Chauhan K, Jandu JS, Brent LH, Al-Dhahir MA. Rheumatoid arthritis. In: StatPearls. Treasure Island, FL: StatPearls Publishing. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK441999> [Last accessed on 2024 Nov 04].
2. Sebbag M, Simon M, Vincent C, Masson-Bessière C, Girbal E, Durieux JJ, *et al.* The antiperinuclear factor and the so-called antikeratin antibodies are the same rheumatoid arthritis-specific autoantibodies. *J Clin Invest* 1995;95:2672-9.
3. Hoet RM, Boerbooms AM, Arends M, Ruiters DJ, van Venrooij WJ. Antiperinuclear factor, a marker autoantibody for rheumatoid arthritis: Colocalisation of the perinuclear factor and profilaggrin. *Ann Rheum Dis* 1991;50:611-8.
4. Schellekens GA, Visser H, de Jong BA, van den Hoogen FH, Hazes JM, Breedveld FC, *et al.* The diagnostic properties of rheumatoid arthritis antibodies recognizing a cyclic citrullinated peptide. *Arthritis Rheum* 2000;43:155-63.
5. Curran AM, Naik P, Giles JT, Darrah E. PAD enzymes in rheumatoid arthritis: Pathogenic effectors and autoimmune targets. *Nat Rev Rheumatol* 2020;16:301-15.
6. Koushik S, Joshi N, Nagaraju S, Mahmood S, Mudeenahally K, Padmavathy R, *et al.* PAD4: Pathophysiology, current therapeutics and future perspective in rheumatoid arthritis. *Expert Opin Ther Targets* 2017;21:433-47.
7. Yang C, Dong ZZ, Zhang J, Teng D, Luo X, Li D, *et al.* Peptidylarginine deiminase 4 as a promising target in drug discovery. *Eur J Med Chem* 2021;226:113840.
8. Yang ML, Sodr  FMC, Mamula MJ, Overbergh L. Citrullination and PAD enzyme biology in type 1 diabetes - regulators of inflammation, autoimmunity, and pathology. *Front Immunol* 2021;12:678953.
9. Wang B, Fields L, Li L. Recent advances in characterization of citrullination and its implication in human disease research: From method development to network integration. *Proteomics* 2023;23:e2200286.
10. Jones JE, Causey CP, Knuckley B, Slack-Noyes JL, Thompson PR. Protein arginine deiminase 4 (PAD4): Current understanding and future therapeutic potential. *Curr Opin Drug Discov Devel* 2009;12:616-27.
11. Bicker KL, Thompson PR. The protein arginine deiminases: Structure, function, inhibition, and disease. *Biopolymers* 2013;99:155-63.
12. Strahl BD, Allis CD. The language of covalent histone modifications. *Nature* 2000;403:41-5.
13. Zee BM, Levin RS, DiMaggio PA, Garcia BA. Global turnover of histone post-translational modifications and variants in human cells. *Epigenetics Chromatin* 2010;3:22.
14. Vossenaar ER, Zendman AJ, van Venrooij WJ, Pruijn GJ. PAD, a growing family of citrullinating enzymes: Genes, features and involvement in disease. *Bioessays*

- 2003;25:1106-18.
15. Thompson PR, Fast W. Histone citrullination by protein arginine deiminase: Is arginine methylation a green light or a roadblock? *ACS Chem Biol* 2006;1:433-41.
  16. Cherrington BD, Morency E, Struble AM, Coonrod SA, Wakshlag JJ. Potential role for peptidylarginine deiminase 2 (PAD2) in citrullination of canine mammary epithelial cell histones. *PLoS One* 2010;5:e11768.
  17. Jang B, Shin HY, Choi JK, Nguyen du PT, Jeong BH, Ishigami A, *et al.* Subcellular localization of peptidylarginine deiminase 2 and citrullinated proteins in brains of scrapie-infected mice: Nuclear localization of PAD2 and membrane fraction-enriched citrullinated proteins. *J Neuropathol Exp Neurol* 2011;70:116-24.
  18. Darrah E, Rosen A, Giles JT, Andrade F. Peptidylarginine deiminase 2, 3 and 4 have distinct specificities against cellular substrates: Novel insights into autoantigen selection in rheumatoid arthritis. *Ann Rheum Dis* 2012;71:92-8.
  19. Aletaha D, Neogi T, Silman AJ, Funovits J, Felson DT, Bingham CO 3<sup>rd</sup>, *et al.* 2010 Rheumatoid arthritis classification criteria: An American College of Rheumatology/European League Against Rheumatism collaborative initiative. *Arthritis Rheum* 2010;62:2569-81.
  20. Sokolove J, Bromberg R, Deane KD, Lahey LJ, Derber LA, Chandra PE, *et al.* Autoantibody epitope spreading in the pre-clinical phase predicts progression to rheumatoid arthritis. *PLoS One* 2012;7:e35296.
  21. Suzuki A, Kochi Y, Shoda H, Seri Y, Fujio K, Sawada T, *et al.* Decreased severity of experimental autoimmune arthritis in peptidylarginine deiminase type 4 knockout mice. *BMC Musculoskelet Disord* 2016;17:205.
  22. Bawadekar M, Shim D, Johnson CJ, Warner TF, Rebernick R, Damgaard D, *et al.* Peptidylarginine deiminase 2 is required for tumor necrosis factor alpha-induced citrullination and arthritis, but not neutrophil extracellular trap formation. *J Autoimmun* 2017;80:39-47.
  23. Willis VC, Banda NK, Cordova KN, Chandra PE, Robinson WH, Cooper DC, *et al.* Protein arginine deiminase 4 inhibition is sufficient for the amelioration of collagen-induced arthritis. *Clin Exp Immunol* 2017;188:263-74.
  24. Willis VC, Gizinski AM, Banda NK, Causey CP, Knuckley B, Cordova KN, *et al.* N- $\alpha$ -benzoyl-N5-(2-chloro-1-iminoethyl)-L-ornithine amide, a protein arginine deiminase inhibitor, reduces the severity of murine collagen-induced arthritis. *J Immunol* 2011;186:4396-404.
  25. Kawalkowska J, Quirke AM, Ghari F, Davis S, Subramanian V, Thompson PR, *et al.* Abrogation of collagen-induced arthritis by a peptidyl arginine deiminase inhibitor is associated with modulation of T cell-mediated immune responses. *Sci Rep* 2016;6:26430.
  26. Shelef MA, Sokolove J, Lahey LJ, Wagner CA, Sackmann EK, Warner TF, *et al.* Peptidylarginine deiminase 4 contributes to tumor necrosis factor  $\alpha$ -induced inflammatory arthritis. *Arthritis Rheumatol* 2014;66:1482-91.
  27. Seri Y, Shoda H, Suzuki A, Matsumoto I, Sumida T, Fujio K, *et al.* Peptidylarginine deiminase type 4 deficiency reduced arthritis severity in a glucose-6-phosphate isomerase-induced arthritis model. *Sci Rep* 2015;5:13041.
  28. Damgaard D, Bawadekar M, Senolt L, Stensballe A, Shelef MA, Nielsen CH. Relative efficiencies of peptidylarginine deiminase 2 and 4 in generating target sites for anti-citrullinated protein antibodies in fibrinogen, alpha-enolase and histone H3. *PLoS One* 2018;13:e0203214.
  29. Sharma M, Damgaard D, Senolt L, Svensson B, Bay-Jensen AC, Nielsen CH, *et al.* Expanding the citrullinome of synovial fibrinogen from rheumatoid arthritis patients. *J Proteomics* 2019;208:103484.
  30. Guo Q, Bedford MT, Fast W. Discovery of peptidylarginine deiminase-4 substrates by protein array: antagonistic citrullination and methylation of human ribosomal protein S2. *Mol Biosyst* 2011;7:2286-95.
  31. Assouhou-Luty C, Rajmakers R, Benckhuijsen WE, Stammen-Vogelzangs J, de Ru A, van Veelen PA, *et al.* The human peptidylarginine deiminases type 2 and type 4 have distinct substrate specificities. *Biochim Biophys Acta* 2014;1844:829-36.
  32. Blachère NE, Parveen S, Frank MO, Dill BD, Molina H, Orange DE. High-titer rheumatoid arthritis antibodies preferentially bind fibrinogen citrullinated by peptidylarginine deiminase 4. *Arthritis Rheumatol* 2017;69:986-95.
  33. Martinez-Prat L, Palterer B, Vitiello G, Parronchi P, Robinson WH, Mahler M. Autoantibodies to protein-arginine deiminase (PAD) 4 in rheumatoid arthritis: Immunological and clinical significance, and potential for precision medicine: Anti-PAD4 antibodies in RA. *Expert Rev Clin Immunol* 2019;15:1073-87.
  34. Ferucci ED, Darrah E, Smolik I, Choromanski TL, Robinson DB, Newkirk MM, *et al.* Prevalence of anti-peptidylarginine deiminase type 4 antibodies in rheumatoid arthritis and unaffected first-degree relatives in indigenous North American Populations. *J Rheumatol* 2013;40:1523-8.
  35. Kan R, Jin M, Subramanian V, Causey CP, Thompson PR, Coonrod SA. Potential role for PADI-mediated histone citrullination in preimplantation development. *BMC Dev Biol* 2012;12:19.
  36. Li P, Li M, Lindberg MR, Kennett MJ, Xiong N, Wang Y. PAD4 is essential for antibacterial innate immunity mediated by neutrophil extracellular traps. *J Exp Med* 2010;207:1853-62.
  37. Li P, Wang D, Yao H, Doret P, Hao G, Shen Q, *et al.* Coordination of PAD4 and HDAC2 in the regulation of p53-target gene expression. *Oncogene* 2010;29:3153-62.
  38. Guo Q, Fast W. Citrullination of inhibitor of growth 4 (ING4) by peptidylarginine deiminase 4 (PAD4) disrupts

- the interaction between ING4 and p53. *J Biol Chem* 2011;286:17069-78.
39. Nakashima K, Arai S, Suzuki A, Nariai Y, Urano T, Nakayama M, *et al.* PAD4 regulates proliferation of multipotent haematopoietic cells by controlling c-myc expression. *Nat Commun* 2013;4:1836.
  40. Martinod K, Witsch T, Erpenbeck L, Savchenko A, Hayashi H, Cherpokova D, *et al.* Peptidylarginine deiminase 4 promotes age-related organ fibrosis. *J Exp Med* 2017;214:439-58.
  41. Caudrillier A, Kessenbrock K, Gilliss BM, Nguyen JX, Marques MB, Monestier M, *et al.* Platelets induce neutrophil extracellular traps in transfusion-related acute lung injury. *J Clin Invest* 2012;122:2661-71.
  42. Thomas GM, Carbo C, Curtis BR, Martinod K, Mazo IB, Schatzberg D, *et al.* Extracellular DNA traps are associated with the pathogenesis of TRALI in humans and mice. *Blood* 2012;119:6335-43.
  43. Cedervall J, Dragomir A, Saupe F, Zhang Y, Ärnlov J, Larsson E, *et al.* Pharmacological targeting of peptidylarginine deiminase 4 prevents cancer-associated kidney injury in mice. *Oncoimmunology* 2017;6:e1320009.
  44. Kolodziej S, Kuvardina ON, Oellerich T, Herglotz J, Backert I, Kohrs N, *et al.* PADI4 acts as a coactivator of Tal1 by counteracting repressive histone arginine methylation. *Nat Commun* 2014;5:3995.
  45. Christophorou MA, Castelo-Branco G, Halley-Stott RP, Oliveira CS, Loos R, Radziszewska A, *et al.* Citrullination regulates pluripotency and histone H1 binding to chromatin. *Nature* 2014;507:104-8.
  46. Xin J, Song X. Role of peptidylarginine deiminase type 4 in gastric cancer. *Exp Ther Med* 2016;12:3155-60.
  47. Chen H, Luo M, Wang X, Liang T, Huang C, Huang C, *et al.* Inhibition of PAD4 enhances radiosensitivity and inhibits aggressive phenotypes of nasopharyngeal carcinoma cells. *Cell Mol Biol Lett* 2021;26:1-12.
  48. Cotechini T, Graham CH. Aberrant maternal inflammation as a cause of pregnancy complications: A potential therapeutic target? *Placenta* 2015;36:960-6.
  49. Venditti CC, Casselman R, Young I, Karumanchi SA, Smith GN. Carbon monoxide prevents hypertension and proteinuria in an adenovirus sFlt-1 preeclampsia-like mouse model. *PLoS One* 2014;9:e106502.
  50. Pang L, Wei Z, Li O, Huang R, Qin J, Chen H, *et al.* An increase in vascular endothelial growth factor (VEGF) and VEGF soluble receptor-1 (sFlt-1) are associated with early recurrent spontaneous abortion. *PLoS One* 2013;8:e75759.
  51. Herraiz I, Dröge LA, Gómez-Montes E, Henrich W, Galindo A, Verlohren S. Characterization of the soluble fms-like tyrosine kinase-1 to placental growth factor ratio in pregnancies complicated by fetal growth restriction. *Obstet Gynecol* 2014;124:265-73.
  52. Andersen LB, Dechend R, Karumanchi SA, Nielsen J, Joergensen JS, Jensen TK, *et al.* Early pregnancy angiogenic markers and spontaneous abortion: An Odense Child Cohort study. *Am J Obstet Gynecol* 2016;215:594.e1-11.
  53. Erpenbeck L, Chowdhury CS, Zsengeller ZK, Gallant M, Burke SD, Cifuni S, *et al.* PAD4 deficiency decreases inflammation and susceptibility to pregnancy loss in a mouse model. *Biol Reprod* 2016;95:132.
  54. Bringmann A, Wiedemann P. Müller glial cells in retinal disease. *Ophthalmologica* 2011;227:1-19.
  55. Wizeman JW, Nicholas AP, Ishigami A, Mohan R. Citrullination of glial intermediate filaments is an early response in retinal injury. *Mol Vis* 2016;22:1137-55.
  56. Wizeman JW, Mohan R. Expression of peptidylarginine deiminase 4 in an alkali injury model of retinal gliosis. *Biochem Biophys Res Commun* 2017;487:134-9.
  57. Rabadi MM, Han SJ, Kim M, D'Agati V, Lee HT. Peptidyl arginine deiminase-4 exacerbates ischemic AKI by finding NEMO. *Am J Physiol Renal Physiol* 2019;316:F1180-90.
  58. Ham A, Rabadi M, Kim M, Brown KM, Ma Z, D'Agati V, *et al.* Peptidyl arginine deiminase-4 activation exacerbates kidney ischemia-reperfusion injury. *Am J Physiol Renal Physiol* 2014;307:F1052-62.
  59. Huang H, Tohme S, Al-Khafaji AB, Tai S, Loughran P, Chen L, *et al.* Damage-associated molecular pattern-activated neutrophil extracellular trap exacerbates sterile inflammatory liver injury. *Hepatology* 2015;62:600-14.
  60. Hakkim A, Fürnrohr BG, Amann K, Laube B, Abed UA, Brinkmann V, *et al.* Impairment of neutrophil extracellular trap degradation is associated with lupus nephritis. *Proc Natl Acad Sci U S A* 2010;107:9813-8.
  61. Knight JS, Subramanian V, O'Dell AA, Yalavarthi S, Zhao W, Smith CK, *et al.* Peptidylarginine deiminase inhibition disrupts NET formation and protects against kidney, skin and vascular disease in lupus-prone MRL/lpr mice. *Ann Rheum Dis* 2015;74:2199-206.
  62. Gordon RA, Herter JM, Rosetti F, Campbell AM, Nishi H, Kashgarian M, *et al.* Lupus and proliferative nephritis are PAD4 independent in murine models. *JCI Insight* 2017;2:92926.
  63. Knight JS, Zhao W, Luo W, Subramanian V, O'Dell AA, Yalavarthi S, *et al.* Peptidylarginine deiminase inhibition is immunomodulatory and vasculoprotective in murine lupus. *J Clin Invest* 2013;123:2981-93.
  64. Villanueva E, Yalavarthi S, Berthier CC, Hodgins JB, Khandpur R, Lin AM, *et al.* Netting neutrophils induce endothelial damage, infiltrate tissues, and expose immunostimulatory molecules in systemic lupus erythematosus. *J Immunol* 2011;187:538-52.
  65. Garcia-Romo GS, Caielli S, Vega B, Connolly J, Allantaz F, Xu Z, *et al.* Netting neutrophils are major inducers of type I IFN production in pediatric systemic lupus erythematosus. *Sci Transl Med* 2011;3:73ra20.
  66. Denny MF, Yalavarthi S, Zhao W, Thacker SG, Anderson M, Sandy AR, *et al.* A distinct subset of proinflammatory neutrophils isolated from patients with systemic lupus erythematosus induces vascular damage and synthesizes type I IFNs. *J Immunol*



- 2010;184:3284-97.
67. Chumanevich AA, Causey CP, Knuckley BA, Jones JE, Poudyal D, Chumanevich AP, *et al.* Suppression of colitis in mice by Cl-amidine: A novel peptidylarginine deiminase inhibitor. *Am J Physiol Gastrointest Liver Physiol* 2011;300:G929-38.
  68. Acharya NK, Nagele EP, Han M, Coretti NJ, DeMarshall C, Kosciuk MC, *et al.* Neuronal PAD4 expression and protein citrullination: Possible role in production of autoantibodies associated with neurodegenerative disease. *J Autoimmun* 2012;38:369-80.
  69. Sokolove J, Brennan MJ, Sharpe O, Lahey LJ, Kao AH, Krishnan E, *et al.* Brief report: Citrullination within the atherosclerotic plaque: A potential target for the anti-citrullinated protein antibody response in rheumatoid arthritis. *Arthritis Rheum* 2013;65:1719-24.
  70. Warnatsch A, Ioannou M, Wang Q, Papayannopoulos V. Inflammation. Neutrophil extracellular traps license macrophages for cytokine production in atherosclerosis. *Science* 2015;349:316-20.
  71. Knight JS, Luo W, O'Dell AA, Yalavarthi S, Zhao W, Subramanian V, *et al.* Peptidylarginine Deiminase inhibition reduces vascular damage and modulates innate immune responses in murine models of atherosclerosis. *Circ Res* 2014;114:947-56.
  72. König MF, Andrade F. A Critical reappraisal of neutrophil extracellular traps and NETosis mimics based on differential requirements for protein citrullination. *Front Immunol* 2016;7:461.
  73. Díaz-Godínez C, Fonseca Z, Néquiz M, Lacleste JP, Rosales C, Carrero JC. Entamoeba histolytica trophozoites induce a rapid non-classical NETosis mechanism independent of NOX2-derived reactive oxygen species and PAD4 activity. *Front Cell Infect Microbiol* 2018;8:184.
  74. Zhou Y, Chen B, Mittereder N, Chaerkady R, Strain M, An LL, *et al.* Spontaneous secretion of the citrullination enzyme PAD2 and cell surface exposure of PAD4 by neutrophils. *Front Immunol* 2017;8:1200.
  75. Khan SA, Edwards BS, Muth A, Thompson PR, Cherrington BD, Navratil AM. GnRH stimulates peptidylarginine deiminase catalyzed histone citrullination in gonadotrope cells. *Mol Endocrinol* 2016;30:1081-91.
  76. van Beers JJ, Zendman AJ, Raijmakers R, Stammen-Vogelzangs J, Pruijn GJ. Peptidylarginine deiminase expression and activity in PAD2 knock-out and PAD4-low mice. *Biochimie* 2013;95:299-308.
  77. Fuchs TA, Brill A, Duerschmied D, Schatzberg D, Monestier M, Myers DD Jr., *et al.* Extracellular DNA traps promote thrombosis. *Proc Natl Acad Sci U S A* 2010;107:15880-5.
  78. Martinod K, Demers M, Fuchs TA, Wong SL, Brill A, Gallant M, *et al.* Neutrophil histone modification by peptidylarginine deiminase 4 is critical for deep vein thrombosis in mice. *Proc Natl Acad Sci U S A* 2013;110:8674-9.
  79. von Brühl ML, Stark K, Steinhart A, Chandraratne S, Konrad I, Lorenz M, *et al.* Monocytes, neutrophils, and platelets cooperate to initiate and propagate venous thrombosis in mice in vivo. *J Exp Med* 2012;209:819-35.
  80. Yu X, Tan J, Diamond SL. Hemodynamic force triggers rapid NETosis within sterile thrombotic occlusions. *J Thromb Haemost* 2018;16:316-29.
  81. Verbij FC, Fijnheer R, Voorberg J, Sorvillo N. Acquired TTP: ADAMTS13 meets the immune system. *Blood Rev* 2014;28:227-34.
  82. Kremer Hovinga JA, Coppo P, Lämmle B, Moake JL, Miyata T, Vanhoorelbeke K. Thrombotic thrombocytopenic purpura. *Nat Rev Dis Primers* 2017;3:17020.
  83. Sorvillo N, Mizurini DM, Coxon C, Martinod K, Tilwawala R, Cherpokova D, *et al.* Plasma peptidylarginine deiminase IV promotes VWF-platelet string formation and accelerates thrombosis after vessel injury. *Circ Res* 2019;125:507-19.
  84. Fadini GP, Menegazzo L, Rigato M, Scattolini V, Poncina N, Bruttocao A, *et al.* NETosis delays diabetic wound healing in mice and humans. *Diabetes* 2016;65:1061-71.
  85. Liang Y, Pan B, Alam HB, Deng Q, Wang Y, Chen E, *et al.* Inhibition of peptidylarginine deiminase alleviates LPS-induced pulmonary dysfunction and improves survival in a mouse model of lethal endotoxemia. *Eur J Pharmacol* 2018;833:432-40.
  86. Zhao X, Gu C, Wang Y. PAD4 selective inhibitor TDFA protects lipopolysaccharide-induced acute lung injury by modulating nuclear p65 localization in epithelial cells. *Int Immunopharmacol* 2020;88:106923.
  87. Martinod K, Fuchs TA, Zitomersky NL, Wong SL, Demers M, Gallant M, *et al.* PAD4-deficiency does not affect bacteremia in polymicrobial sepsis and ameliorates endotoxemic shock. *Blood* 2015;125:1948-56.
  88. Costa NA, Gut AL, Azevedo PS, Polegato BF, Magalhães ES, Ishikawa LLW, *et al.* Peptidylarginine deiminase 4 concentration, but not PAD4 polymorphisms, is associated with ICU mortality in septic shock patients. *J Cell Mol Med* 2018;22:4732-7.
  89. Sun B, Dwivedi N, Bechtel TJ, Paulsen JL, Muth A, Bawadekar M, *et al.* Citrullination of NF- $\kappa$ B p65 promotes its nuclear localization and TLR-induced expression of IL-1 $\beta$  and TNF $\alpha$ . *Sci Immunol* 2017;2:eal3062.
  90. Hanata N, Shoda H, Hatano H, Nagafuchi Y, Komai T, Okamura T, *et al.* Peptidylarginine Deiminase 4 promotes the renal infiltration of neutrophils and exacerbates the TLR7 agonist-induced lupus mice. *Front Immunol* 2020;11:1095.
  91. Cela D, Knackstedt SL, Groves S, Rice CM, Kwon JT, Mordmüller B, *et al.* PAD4 controls chemoattractant production and neutrophil trafficking in malaria. *J Leukoc Biol* 2022;111:1235-42.
  92. Li T, Peng R, Wang F, Hua L, Liu S, Han Z, *et al.* Lysophosphatidic acid promotes thrombus stability by

- inducing rapid formation of neutrophil extracellular traps: A new mechanism of thrombosis. *J Thromb Haemost* 2020;18:1952-64.
93. Yadav R, Yoo DG, Kahlenberg JM, Bridges SL Jr, Oni O, Huang H, *et al.* Systemic levels of anti-PAD4 autoantibodies correlate with airway obstruction in cystic fibrosis. *J Cyst Fibros* 2019;18:636-45.
  94. Rohrbach AS, Slade DJ, Thompson PR, Mowen KA. Activation of PAD4 in NET formation. *Front Immunol* 2012;3:360.
  95. Rodríguez-Rodríguez M, Herrera-Esparza R, Bollain Y Goytia JJ, Pérez-Pérez ME, Pacheco-Tovar D, Murillo-Vázquez J, *et al.* Activation of peptidylarginine deiminase in the salivary glands of Balb/c mice drives the citrullination of Ro and La ribonucleoproteins. *J Immunol Res* 2017;2017:8959687.
  96. Stobernack T, du Teil Espina M, Mulder LM, Palma Medina LM, Piebenga DR, Gabarrini G, *et al.* A Secreted bacterial peptidylarginine deiminase can neutralize human innate immune defenses. *mBio* 2018;9:e01704-18.
  97. Sabnis RW. Novel peptidylarginine deiminase Type 4 (PAD4) inhibitors. *ACS Med Chem Lett* 2022;13:1537-8.
  98. Martín Monreal MT, Rebak AS, Massarenti L, Mondal S, Šenolt L, Ødum N, *et al.* Applicability of small-molecule inhibitors in the study of peptidyl arginine deiminase 2 (PAD2) and PAD4. *Front Immunol* 2021;12:716250.
  99. Naik P, Shi J, Andrade F, Darrah E. Antibodies to PAD4 Drive Monocyte Activation and Differentiation Into Osteoclast-Like Cells. Available from: <https://acrabstracts.org/abstract/antibodies-to-pad4-drive-monocyte-activation-and-differentiation-into-osteoclast-like-cells> [Last accessed on 2024 Nov 04].
  100. Krishnamurthy A, Ytterberg AJ, Sun M, Sakuraba K, Steen J, Joshua V, *et al.* Citrullination controls dendritic cell transdifferentiation into osteoclasts. *J Immunol* 2019;202:3143-50.
  101. Willemze A, Trouw LA, Toes RE, Huizinga TW. The influence of ACPA status and characteristics on the course of RA. *Nat Rev Rheumatol* 2012;8:144-52.
  102. Ellebrecht CT, Bhoj VG, Nace A, Choi EJ, Mao X, Cho MJ, *et al.* Reengineering chimeric antigen receptor T cells for targeted therapy of autoimmune disease. *Science* 2016;353:179-84.
  103. Benham H, Nel HJ, Law SC, Mehdi AM, Street S, Ramnarth N, *et al.* Citrullinated peptide dendritic cell immunotherapy in HLA risk genotype-positive rheumatoid arthritis patients. *Sci Transl Med* 2015;7:290ra87.
  104. Liu X, Arfman T, Wichapong K, Reutelingsperger CPM, Voorberg J, Nicolaes GA. PAD4 takes charge during neutrophil activation: Impact of PAD4 mediated NET formation on immune-mediated disease. *J Thromb Haemost* 2021;19:1607-17.
  105. Thanabalasuriar A, Scott BNV, Peiseler M, Willson ME, Zeng Z, Warrener P, *et al.* Neutrophil extracellular traps confine *Pseudomonas aeruginosa* ocular biofilms and restrict brain invasion. *Cell Host Microbe* 2019;25:526-36.e4.
  106. Lu Y, Peng Z, Zhu D, Jia Y, Taledaohan A, Li Y, *et al.* RGD Peptide and PAD4 inhibitor-loaded gold nanorods for chemo-photothermal combined therapy to inhibit tumor growth, prevent lung metastasis and improve biosafety. *Int J Nanomedicine* 2021;16:5565-80.
  107. Gajendran C, Fukui S, Sadhu NM, Zainuddin M, Rajagopal S, Gosu R, *et al.* Alleviation of arthritis through prevention of neutrophil extracellular traps by an orally available inhibitor of protein arginine deiminase 4. *Sci Rep* 2023;13:3189.