

Ethosomes as Promising Novel Transdermal Drug Delivery System and their Therapeutic Applications – A Review

Jyothi Mahadev^{1,2}, S. Nirmala³

¹Department of Pharmacy, Bharath Institute of Higher Education and Research, Chennai, Tamil Nadu, India,

²Department of Pharmaceutics, East Point College of Pharmacy, Jnana Prabha, Bengaluru, Karnataka, India,

³Department of Pharmacognosy, Faculty of Pharmacy, Sree Balaji Medical College and Hospital Campus, Bharath Institute of Higher Education and Research, Chennai, Tamil Nadu, India

Abstract

Inadequate penetration of therapeutically active chemicals is the main factor affecting transdermal drug delivery system networks. The stratum corneum (SC) serves as the skin's initial line of defense against drug molecule penetration. To date, this field has employed a number of tactics to overcome the SC barrier limits. The main purpose of a few innovative methods, including as liposomes, niosomes, ethosomes, and transfersomes, is to make pharmaceutical and cosmetic substances more permeable through the SC barrier. The melting point of SC lipids is lowered by ethosomes, which are stable, elastic, nanoscale vesicles with high phospholipid and ethanol content that interact with the polar head domain of lipid molecules. Finally, it increases lipid fluidity and cell membrane permeability. This section provides an overview of the most recent ethosome updates, depending on several pharmacological dose forms, involving gels, patches, and creams. There is also a brief discussion on updated ethosome patents. To sum up, ethosomes are an ideal vehicle for the delivery of drugs, cosmetics, and other substances and can be used in place of traditional pharmaceutical applications.

Key words: Anti-psoriatic, ethosomes, liposomes, stratum corneum, transfersomes

INTRODUCTION

The most versatile and broad route for topical and systemic drug administration is the skin. An approach to pharmaceutical administration that is less invasive is transdermal drug delivery (TDD), which provides less frequent dosage, controlled drug distribution, patient compliance, as well as prevention of first-pass metabolism.^[1-4] Following transfersomes, groundbreaking research led to the identification of ethosomes, a distinct lipid vesicular system.^[5] Ethosomes are novel modified lipid carriers composed of phospholipids, water, and ethanol. In addition to phospholipids and water, ethosomes contain comparatively high amounts of ethanol, which may have enhanced vesicular properties and epidermal penetration.^[6-10]

WORKING MECHANISM OF ETHOSOMES

In the activity of ethosomes, ethanol, vesicles, and skin lipids work in concert.^[11] The dispersion of active substances is improved over liposomes by ethosomes and skin lipids because they interact more effectively. In addition, ethanol renders vesicles flexible and smooth, allowing for deeper penetration into the epidermal layer.^[12]

Address for correspondence:

S. Nirmala, Department of Pharmacognosy, Faculty of Pharmacy, Sree Balaji Medical College and Hospital Campus, Bharath Institute of Higher Education and Research, Chromepet, Chennai - 600044, Tamil Nadu, India.
E-mail: jyothi.epcp@gmail.com

Received: 15-04-2025

Revised: 13-06-2025

Accepted: 25-06-2025

Table 1

Ethosomal system types

S.no	Parameter	Classical E thosomes	Binary E thosomes	Transethosomes
1	Composition	Phospholipids, ethanol and stabilizer	Phospholipids, ethanol stabilizer and propylene glycol	Phospholipids, ethanol, edge activator and charge inducer
2	Morphology	Spherical	Spherical	Spherical
3	Size	Smaller than classical ethosomes	Equal or Smaller than classical ethosomes	Size varies based on change of the inducer concentration
4	Entrapment efficiency	Greater than traditional ethosomes	Often Greater than traditional ethosomes	Higher than majority of ethosomes
5	Skin permeation	High	High	High
6	Stability	Highly robust	Stable	No evidence

ETHOSOMAL SYSTEM TYPES

Ethosomal systems, used for transdermal drug delivery, are generally categorized into three types based on their composition: classical ethosomes, binary ethosomes, and transethosomes. These systems are characterized by the presence of ethanol, phospholipids, and water (Table 1 and Figure 1).

MECHANISM OF PENETRATION

Ethanol decreases “transition temperature of lipids in the stratum corneum (SC), increases their fluidity, and decreases the density of lipid multilayers through interacting with lipid molecules in the polar head group region. “Ethosome” effect” follows, whereby the medicine is released into deep layers of skin by lipid penetration as well as permeation through the creation of new routes as a result of the ethosomes’ fusing and malleability with skin lipids.^[13,14]

RATIONALE FOR TDD

In other instances, the most practicable method of drug ingestion – the oral route – was impractical; therefore, other methods had to be looked for. TDD has numerous obvious advantages, including convenience of administration, ease of stopping therapy, and a relatively large as well as easily accessible surface area for absorption.^[15,16]

COMPOSITION OF ETHOSOMES

Although ethersomes and liposomes both have a lipid bilayer, their composition (high ethanol concentration) sets them apart. Hydroalcoholic or glycolic/hydrophospholipids that make up ethosomes have comparatively high alcohol

content. Between 22% and 70% of the alcohol and glycol mixture may be present in the non-aqueous phase.^[17]

PREPARATION OF ETHOSOMES

Ethosomes can be prepared in a variety of ways, which might change based on the particular circumstances and needs of the process. They are a good substitute for conventional medication delivery methods because of their rather easy and affordable manufacture.^[18]

Table 2 highlights “the most important methods that can be used to prepare ethosomes.

ADVANTAGE OF ETHOSOMAL DRUG DELIVERY SYSTEM

Comparing ethosomal drug delivery systems to other transdermal and dermal administration techniques reveals several advantages. The ability to distribute large molecules (like protein or peptide molecules), use of non-toxic raw ingredients in formulation, and enhanced drug penetration through the skin for transdermal drug administration are some advantages.

DISADVANTAGES OF ETHOSOMAL DRUG DELIVERY

Precipitation might result from ethosomes with poor shelling grouping together. Sufficient medication solubility conditions are required to enter the cutaneous microcirculation and systemic circulation. When transferred into water, the ethosomes may aggregate and break down if shell locking is insufficient.^[19-25]

EVALUATION OF ETHOSOMES^[26]

Vesicle-skin interaction study by transmission electron microscopy (TEM) and scanning electron microscopy (SEM)

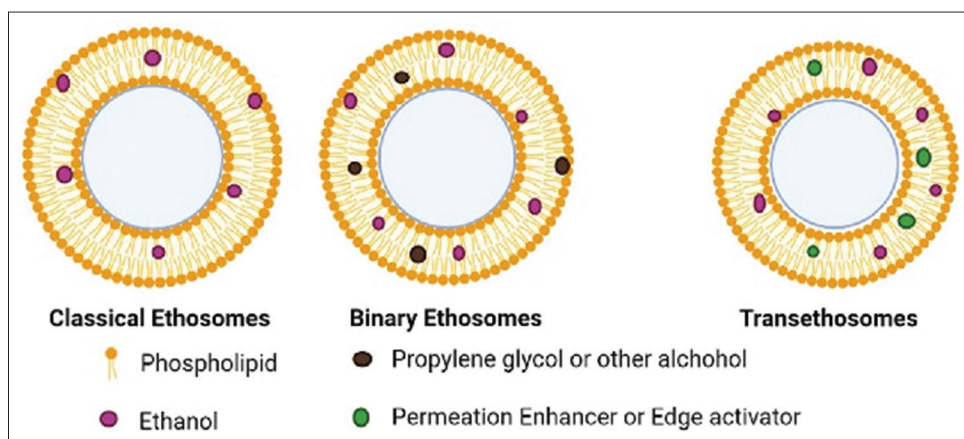
By “interacting with lipid molecules in the polar head group region, ethanol lowers the transition temperature of lipids in the SC, enhances their fluidity, and reduces the density of lipid multilayers.

Filter membrane-vesicle interaction study by SEM

Before being placed in diffusion cells, filter membrane with 50 nm pore size was coated with vesicle suspension (0.2 mL). Phosphate buffer saline solution (PBS), which has a pH of

Table 2: Preparation of ethosomes with key characteristics

Method	Advantages	Cost	Manufacturing scale
Hot method	<ul style="list-style-type: none"> • Quick and easy method • Smaller particle size • Stable at RT • Suitable for hydrophilic and hydrophobic drugs • High encapsulation efficiency 	Low to moderate	Small to large
Cold method	<ul style="list-style-type: none"> • Simple and mild preparation method • Loading capacity is good • Stable at RT • Suitable for hydrophilic and hydrophobic drugs • Scalability is good 	Low to moderate	Small to large
Vortex method	<ul style="list-style-type: none"> • Encapsulation capacity is high • Particle size is small • Loading capacity is good • Stable at RT • Suitable for hydrophilic and hydrophobic drugs 	Moderate to high	Small to moderate
Rotary film evaporation	<ul style="list-style-type: none"> • Loading capacity is high • Stable • Suitable only for hydrophobic drugs 	Moderate to high	Small to moderate
Classic dispersion method	<ul style="list-style-type: none"> • Simple and mild preparation method • Suitable for hydrophilic and hydrophobic drugs 	Low to moderate	Small to large
Transmembrane pH-gradient method	<ul style="list-style-type: none"> • Stable • Suitable for only hydrophobic drugs 	High	Small to moderate

**Figure 1:** Pictorial representation of ethosomes

6.5, came into contact with the lower side of the filter while the upper side has been left open to air.

Vesicle-skin interaction study by fluorescence microscopy

The fluorescence microscopy has been examined using TEM and SEM techniques. Five μm -thick pieces of paraffin blocks that had been cut with “microtome (Erma Optical Works, Tokyo, Japan) have been examined under fluorescence microscope. MT-2 Cell” Cytotoxicity Test.

STABILITY OF ETHOSOMES

Entrapment capacity and particle size have been used to assess the compositions’ long-term stability. The entrapment efficiency is improved by raising the ethanol level from 15% to 45% because the membranes are more flexible. The vesicle membrane may become more porous at ethanol concentrations above 45%, which would reduce the efficiency of entrapment. As a result, the ethosomes become unstable. Lipid hydrolysis produces lyso-PC. Because lyso-PC enhances ethosome permeability, it is crucial to keep a minimum in a given preparation.^[27-29]

ETHOSOMES INCORPORATED IN SUITABLE DOSAGE FORM

Before being placed in diffusion cells, filter membrane with 50 nm pore size was coated with vesicle suspension (0.2 mL). Upper portion of filter has been exposed to air, while lower portion has been in contact with PBS, which has a pH of 6.5.

ETHOSOMES IN GEL FORM

Ethosomal gels are distinguished by their pH, viscosity, extrudability, and spreadability. In all of their related categories, carbopol and hydroxypropyl methylcellulose are the two most widely utilized gel-forming agents for integrating ethosomal systems.^[30-35]

APPLICATION OF ETHOSOMES IN DRUG DELIVERY

In recent two decades, ethosomes were widely utilized for transdermal/topical drug delivery applications. The plentiful literature survey suggested that the novel ethosomes have to gain immense importance in advanced drug delivery systems, especially delivery through skin membranes.

ANTIFUNGAL AGENT DELIVERY

In the lipid carrier system, ethosomes show an incredible ability to enhance transdermal permeation. Ethosomes-based formulations have been designed to achieve a rapid onset of action, maximum drug release, and fewer side effects. There is no harmful effect to the skin during drug transport to systemic circulation across undamaged skin. Furthermore, it reduces the duration of treatment.^[36,37] The *in vivo* research confirmed that drug-loaded ethosomes were more active and effective against *Candida parapsilosis* than *Aspergillus niger* due to the synergistic effect from need and drug. Therefore, it can be used as a potential antifungal agent in biomedical applications.^[38,39] Finally, ethosomes have the potential to be a vehicle for topical administration of fluconazole.^[40-42]

ANTI-INFLAMMATORY DRUG

From its inception, an anti-inflammatory agent involving non-steroidal anti-inflammatory drugs (NSAIDs) is generally utilized for the treatment of chronic pain and inflammation. When taking NSAIDs orally, one may experience stomach ulcers, abdominal pain, irregularities, and various vascular problems. In addition, NSAIDs can fail a quick onset of action owing to their low bioavailability and half-life.^[43-50] Furthermore, the paeonol-ethosomes *in vitro* skin penetration

and skin retention study confirmed the high permeation rate (138.58 $\mu\text{g}/\text{cm}^2$) and skin retention (52.60 $\mu\text{g}/\text{cm}^2$). In nutshell, ethosomes could be an exceptional nanocarrier for potential drug delivery applications, especially transdermal administration of paeonol.^[51,52] The ethanol offers softness and flexibility to the ethosomal vesicles, which results in high penetration. As well, the high concentration of ethanol increased the skin permeation rate.^[53,54]

ANTI-VIRAL DRUGS

Acyclovir-loaded ethosomal gel has been reported for the management of Herpes zoster. It has been prepared using ethanol, phospholipid, and PEG through the cold method. The optimized batch showed about -20.5 mV zeta potential and about 331.69 nm vesicle size. Finally, 1% w/w carbopol 980 showed better *in vitro* acyclovir drug release (82.23%) after 8 h, and release kinetics was found in the zero-order model ($R^2: 0.989$). Therefore, it can be used as a budding vehicle for the delivery of the antiviral agent for an extended time in the case of the affected skin part.^[55,56]

ETHOSOMES IN COSMECEUTICALS

Various cosmetic preparations contain numerous active ingredients that only help to offer appropriate penetration to the SC layer of the skin. However, a few of the formulations used topically have relatively lower potency because the SC is resistant to being transported into the skin. For this reason, the formulation needs to be modified. Accordingly, the appropriate modification in the formulation will lead to the high permeability of the formulation containing a drug. Interestingly, ethosomes can be used as an excellent carrier to supply a wide range of ingredients through topical routes.^[57,58]

ANTICANCER DRUG DELIVERY

In the world of medical sciences, cancer therapy is still a challenging task. It may be due to the inefficiency of the existing clinical approaches used for the treatment, which results in a large number of deaths occurring each year. Hence, there is an urge to expand a suitable carrier for the delivery of the drug to the targeted area and systemic application.^[59-61]

DELIVERY OF ANALGESIC AGENT

Nowadays, effective pain management is a challenging task for researchers. It may be because of severe side effects of the different analgesic agents.^[62]

DELIVERY OF ANTI-PSORIATIC AGENT

Psoriasis is a chronically non-infectious skin condition, joint, and/or both with relapsing inflammatory and hyperkeratotic plaque episodes. In such conditions, efficient delivery of the drug is an important factor during formulation design.^[63]

DELIVERY OF ANTI-HYPERTENSIVE AGENT

Over the past decades, hypertension incidence has increased dramatically. Moreover, it is also the leading cause of death also. For the last couple of decades, the scientific fraternity has been working on various effective carriers for the delivery of anti-hypertensive agents through different routes.

CONCLUSION

Ethosomes have demonstrated encouraging outcomes in the pharmaceutical industry as drug delivery systems, and research into their potential for skin care applications is growing. Targeted drug delivery, improved drug-loading capacity, and higher stability are made possible by the inclusion of ethanol in their composition.

ACKNOWLEDGMENT

For their unwavering support and encouragement in helping me write this review article, I am extremely grateful to my research supervisor at the “Bharath Institute of Higher Education and Research in Selaiyur, Chennai, Tamil” Nadu, India.

REFERENCES

1. Prausnitz MR, Mitragotri S, Langer R. Current status and future potential of transdermal drug delivery. *Nat Rev Drug Discov* 2004;3:115-24.
2. Prausnitz MR, Langer R. Transdermal drug delivery. *Nat Biotechnol* 2008;26:1261-8.
3. Schoellhammer CM, Blankschtein D, Langer R. Skin permeabilization for transdermal drug delivery: Recent advances and future prospects. *Expert Opin Drug Deliv* 2014;11:393-407.
4. Mezei M, Gulasekharan V. Liposomes -- a selective drug delivery system for the topical route of administration I. Lotion dosage form. *Life Sci* 1980;26:1473-7.
5. Tuitou E. Composition for Applying Active Substances to or through the Skin, US Patents no 5:716. p. 638.
6. Tuitou E, Dayan N, Bergelson L, Godin B, Eliaz M. Ethosomes - novel vesicular carriers for enhanced delivery: Characterization and skin penetration properties. *J Contr Release* 2000;65:403-18.
7. Ainbinder D, Tuitou E. A new approach for skin tumor treatment: From delivery system characterization to *in vivo* evaluation. *Drug Deliv Transl Res* 2011;1:53-65.
8. Jain S, Patel N, Madan P, Lin S. Quality by design approach for formulation, evaluation and statistical optimization of diclofenac-loaded ethosomes via transdermal route. *Pharm Dev Technol* 2015;20:473-89.
9. Sarwa K, Suresh P, Rudrapal M, Verma V. Penetration of tamoxifen citrate loaded ethosomes and liposomes across human skin: A comparative study with confocal laser scanning microscopy. *Curr Drug Deliv* 2014;11:332-7.
10. Cevc G, Blume G. Lipid vesicles penetrate into intact skin owing to the transdermal osmotic gradients and hydration force. *Biochim Biophys Acta* 1992;1104:226-32.
11. Abdulbaqi LM, Darwis Y, Khan NA, Khan AA. Ethosomal nanocarriers: The impact of constituents and formulation techniques on ethosomal properties, *in vivo* studies, and clinical trials. *Int J Nanomed* 2016;11:2279-2304.
12. Ainbinder D, Tuitou E. Testosterone ethosomes for enhanced transdermal delivery. *Drug Deliv J Deliv Target Ther Agents* 2005;12:297-303.
13. Verma DD, Fahr A. Synergistic penetration enhancement effect of ethanol and phospholipids on the topical delivery of cyclosporin A. *J Control Release* 2004;97:55-66.
14. Dubey V, Mishra D, Jain NK. Melatonin loaded ethanolic liposomes: Physicochemical characterization and enhanced transdermal delivery. *Eur J Pharm Biopharm* 2007;67:398-405.
15. Guy RH. Ethosomes an recent approach in transdermal drug delivery system. *Int J Pharm* 1985;6:112-6.
16. Panchagnula R, Pillai O, Nair VB, Ramarao P. Transdermal iontophoresis revisited. *Curr Opin Chem Bio* 2000;4:468-73.
17. Mustafa MA. Deformable Liposomes and Ethosomes: Mechanism of Enhanced Skin Delivery. *Int J Pharm* 2006;322:60-6.
18. Ahad A, Akhtar N, Gupta DK. Ethosomes: A potential vesicular carrier for drug delivery. In: *Systems of Nanovesicular Drug Delivery*. United States: Academic Press; 2022. p. 221-37.
19. Jain H, Patel J, Joshi K, Patel P, Upadhyay UM. Ethosomes: A novel drug carrier. *Int J Clin Pract* 2011;7:1-4.
20. Upadhyay N, Mandal S, Bhatia L, Shailesh S, Chauhan P. A review on ethosomes: An emerging approach for drug delivery through the skin. *Recent Res Sci Technol* 2011;3:19-24.
21. Sivakranth M, Ara PA, Krishnaveni C, Venkatesh E. Ethosomes: A novel vesicular drug delivery system. *Int J Adv Pharmaceu Res* 2012;2:16-27.
22. Kumar R, Aslam MD, Tripathi A, Prasad D, Chaudhary V, Jain V, *et al.* Ethosomes: Novel vesicular carriers in transdermal drug delivery. *J Global Pharma Technol* 2010;2:1-7.

23. Rathore AR, Khambete H, Jain S. Preparation and characterization of repaglinide loaded ethosomal gel for the treatment of NIDDM. *Int J Pharm Biol Arch* 2013;4:385-90.
24. Shahwal V, Samnani A, Dubey B, Bhowmick M. Ethosomes: An overview. *Int J Biomed Adv Res* 2011;2:161-8.
25. Dhurve R, Kashyap N, Mishra A, Pathak AK. A holistic review on ethosome: A promising drug delivery system for topical fungal disease. *Int J Pharm Biol Arch* 2014;5:13-26.
26. Ghule AR, Shinkar DM, Saudagar RB. Ethosomes: Carrier for enhanced transdermal drug delivery system. *J Adv Pharm Edu Res* 2014;4:380-7.
27. Patel A, Sharma R, Trivedi M, Panicker A. Ethosomes: A novel tool for transdermal drug delivery. *Res J Pharm Tech* 2013;6:838-41.
28. Touitou E, Godin B. Ethosomes for skin delivery. *J Drug Deliv Sci Technol* 2007;17:303-8.
29. Rao LS. Preparation of liposomes on the industrial scale: Problems and perspectives. *Liposome Technol* 2019;14:247-57.
30. Fu X, Shi Y, Wang H, Zhao X, Sun Q, Huang Y. Ethosomal gel for improving transdermal delivery of thymosin β -4. *Int J Nanomedicine* 2019;14:9275-84.
31. Ismail TA, Shehata TM, Mohamed DI, Elsewedy HS, Soliman WE. Quality by design for development, optimization and characterization of Brucineethosomal gel for skin cancer delivery. *Molecules* 2021;26:3454.
32. Saraf S, Kumar Gupta M. Itraconazole loaded ethosomal gel system for efficient treatment of skin cancer. *Int J Drug Deliv* 2018;10:12-9.
33. Khan NR, Wong TW. Microwave-aided skin drug penetration and retention of 5-fluorouracil-loaded ethosomes. *Expert Opin Drug Deliv* 2016;13:1209-19.
34. Gollavilli H, Hegde AR, Managuli RS, Bhaskar KV, Dengale SJ, Reddy MS. Naringinnano-ethosomal novel sunscreen creams: Development and performance evaluation. *Colloids Surf B Biointerfaces* 2020;193:111122.
35. Song X, Jiang Y, Zhang W, Elfawal G, Wang K, Jiang D. Transcutaneous tumor vaccination combined with anti-programmed death-1 monoclonal antibody treatment produces a synergistic antitumor effect. *Acta Biomater* 2022;140:247-60.
36. Saratchandarn C, Shirwaikar A, Devi AS, Vipin K. Comparative evaluation of sonicated and un-sonicated ethosomes containing ketoconazole. *Int J Adv Pharm Biol Chem* 2012;1:15-20.
37. Johnsen S, Bennett E, Jensen VG. Therapeutic effectiveness of oral testosterone. *Lancet* 1974; 304:1473-5.
38. Dave V, Bhardwaj N, Gupta N, Tak K. Herbal ethosomal gel containing luliconazole for productive relevance in the field of biomedicine. *Biotech* 2020;10:1-15.
39. Begum SB, Kishor DV, Begum NU. Formulation and *in-vitro* evaluation of ciclopiroxethosomal gel. *J Drug Dev Deliv* 2019;2:8-16.
40. Dhurve R, Mishra A. Formulation and evaluation of ethosomal gel of fluconazole for topical drug delivery. *Int J Curr Trends Drug Dev Indust Pharm* 2019;3:47.
41. Kumar JR. Anticandidal activity of ethosomal gel containing miconazole nitrate in male Sprague Dawley rat. *J Pharm Sci Res* 2018;10:3400-5.
42. Mbah CC, Builders PF, Agubata CO, Attama AA. Development of ethosomal vesicular carrier for topical application of griseofulvin: Effect of ethanol concentration. *J Pharm Investig* 2019;49:27-36.
43. Nangare S, Jadhav N, Ghagare P, Muthane T. Pharmaceutical applications of electrospinning. *Ann Pharm Fr* 2020;78:1-11.
44. Yamazaki R, Kawai S, Matsuzaki T, Kaneda N, Hashimoto S, Yokokura T. Aceclofenac blocks prostaglandin E2 production following its intracellular conversion into cyclooxygenase inhibitors. *Eur J Pharmacol* 1997;329:181-7.
45. Barupal A, Gupta V, Ramteke S. Preparation and characterization of ethosomes for topical delivery of aceclofenac. *Indian J Pharm Sci* 2010;72:582-6.
46. Sujitha B, Krishnamoorthy B, Muthukumaran M. Formulation and evaluation of piroxicam loaded ethosomal gel for transdermal delivery. *Int J Adv Pharm Gen Res* 2014;2:34-45.
47. Garg V, Singh H, Bhatia A, Raza K, Singh SK, Beg S. Systematic development of transethosomal gel system of piroxicam: Formulation optimization, *in vitro* evaluation, and *ex vivo* assessment. *AAPS Pharm Sci Tech* 2017;18:58-71.
48. Paliwal S, Tilak A, Sharma J, Dave V, Sharma S, Yadav R. Flurbiprofen loaded ethosomes-transdermal delivery of anti-inflammatory effect in rat model. *Lipids Health Dis* 2019;18:1064.
49. Supraja R, Sailaja AK. Formulation of mefenamic acid loaded ethosomal gel by hot and cold methods. *Nano Biomed Eng* 2017;9:27-35.
50. Sakdiset P, Amnuait T, Pichayakorn W, Pinsuwan S. Formulation development of ethosomes containing indomethacin for transdermal delivery. *J Drug Deliv Sci Technol* 2019;52:760-8.
51. Ma H, Guo D, Fan Y, Wang J, Cheng J, Zhang X. Paeonol-loaded ethosomes as transdermal delivery carriers: Design, preparation and evaluation. *Molecules* 2018;23:1756.
52. Chowdary VS, Azmeera SM, Begum AM, Sai SR, Vishwanath N. Design, development and evaluation of celecoxib loaded ethosomal gel. *J Pharma Creat* 2017;4:45-54.
53. Ghanbarzadeh S, Arami S. Enhanced transdermal delivery of diclofenac sodium via conventional liposomes, ethosomes, and transfersomes. *Biomed Res Int* 2013;2013:616810.
54. Mistry A, Ravikumar P. Development and evaluation of azelaic acid based ethosomes for topical delivery for the treatment of acne. *Indian J Pharm Educ Res*

- 2016;50:S232-43.
55. Shukla KV, Sharma A, Yadav M. Formulation development and evaluation of ethosomal gel of acyclovir for the treatment of herpes zoster. *J Drug Deliv Ther* 2019;9:664-8.
56. Gupta AB, Jain AP, Gupta S. Formulation and evaluation of Lamivudine ethosomes for the treatment of AIDS disease. *J Emerg Technol Innov Res* 2019;6:271-84.
57. Madsen JT, Vogel S, Karlberg AT, Simonsson C, Johansen JD, Andersen KE. Ethosome formulations of known contact allergens can increase their sensitizing capacity. *Acta Derm Venereol* 2010;90:374-8.
58. Jankowski A, Dyja R, Sarecka-Hujar B. Dermal and transdermal delivery of active substances from semisolid bases. *Indian J Pharm Sci* 2017;79:488-500.
59. Verma P, Pathak K. Therapeutic and cosmeceutical potential of ethosomes: An overview. *J Adv Pharm Technol Res* 2010;1:274-82.
60. Cristiano MC, Froiio F, Spaccapelo R, Mancuso A, Nisticò SP, Udongo BP. Sulforaphane-loaded ultradeformable vesicles as a potential natural nanomedicine for the treatment of skin cancer diseases. *Pharmaceutics* 2020;12:6.
61. Peram MR, Jalalpure S, Kumbar V, Patil S, Joshi S, Bhat K. Factorial design based curcumin ethosomal nanocarriers for the skin cancer delivery: *In vitro* evaluation. *J Liposome Res* 2019;29:291-311.
62. Sundar VD, Divya P, Dhanaraju MD. Design development and characterization of tramadol hydrochloride loaded ethosomes. *Acta Derm Venereol* 2021;90:374-8.
63. Shelke O, Kulkarni A. Formulation, development and evaluation of nanoethosomal gel of tramadol hydrochloride. *J Nanomed Nanotechnol* 2018;9:1000514.

Source of Support: Nil. **Conflicts of Interest:** None declared.