

Green Synthesis of Selenium Oxide Nanoparticles Using *Pleurotus ostreatus* (Oyster Mushroom) Preparation Characterization and Antimicrobial Efficacy

Saeed Anawalikar, S. Delphine Priscilla Antony, Sanyukta Singh

Department of Conservative Dentistry and Endodontics, Saveetha Dental College and Hospitals, Saveetha Institute of Medical and Technical Sciences, Saveetha University, Chennai Tamil Nadu, India

Abstract

This study aimed to environmentally synthesize selenium oxide nanoparticles (SeO₂ NPs) using *Pleurotus ostreatus* (oyster mushroom) extract and evaluated their antimicrobial efficacy against *Streptococcus mutans* and *Enterococcus faecalis*. SeO₂ NPs were synthesized via an aqueous extract of *P. ostreatus* and characterized using ultraviolet-visible spectroscopy, Fourier transform infrared spectroscopy, and X-ray diffraction (XRD) to determine their optical, functional, and structural properties. The antimicrobial efficacy of the synthesized nanoparticles was assessed against oral pathogens *S. mutans* and *E. faecalis* using the agar well diffusion method. Scanning electron microscopy (SEM) was used to analyze SeO₂ NPs surface shape and size distribution. SEM scans showed that SeO₂ NPs are homogeneous and spherical with an average particle size of up to 10 nm. Well-dispersed particles show less aggregation. The XRD pattern shows no extra peaks, indicating that the synthesized nanoparticles are phase-pure. SeO₂ NPs' antibacterial activity increases with concentration. Positive control (PC), a traditional antibiotic, has the largest inhibitory zones for both bacteria, proving its efficacy. At a dosage of 100 µg/mL, SeO₂ NPs approach the PC's efficiency, suggesting potential as an alternative antibacterial agent. SeO₂ NPs were synthesized utilizing a green method and characterized using XRD and antibacterial tests. The major findings reveal that green-chemistry SeO₂ NPs are efficient against *S. mutans* and *E. faecalis*. Nanoparticles were effective at various doses, with the strongest inhibitory zones at higher ones. Green synthesis is necessary to avoid dangerous chemicals and protect the environment.

Key words: Antibacterial activity, biocompatibility, *Enterococcus faecalis*, green synthesis, selenium oxide nanoparticles, *Streptococcus mutans*, X-ray diffraction

INTRODUCTION

Nanomaterials have emerged as promising antibacterial agents owing to their unique physicochemical properties, including ultra-small particle size, high surface-area-to-volume ratio, and enhanced chemical reactivity.^[1] These characteristics enable nanomaterials to effectively interact with microbial cells and disrupt their integrity, positioning them as innovative tools in combating infections.^[2] Nanomaterials, particularly nanoparticles, are playing an increasingly important role in modern oral healthcare. In the management of dental infections, nanoparticles exhibit potent antibacterial and antibiofilm properties. Their

ability to disintegrate bacterial membranes, form reactive oxygen species (ROS), and impede microbial metabolism makes them effective against strains like *Enterococcus faecalis* and *Streptococcus mutans*, which are often associated with persistent endodontic infections and dental caries.^[3]

Address for correspondence:

S. Delphine Priscilla Antony, Department of Conservative Dentistry and Endodontics, Saveetha Dental College, Saveetha Institute of Medical and Technical Sciences, Saveetha University, Chennai, Tamil Nadu, India.

E mail: delphy.priscilla@gmail.com

Received: 15-08-2025

Revised: 24-09-2025

Accepted: 30-09-2025

Nanoparticles have also been explored for use in dental restorative materials, such as nanocomposites, where they improve mechanical strength, wear resistance, and antimicrobial performance. In preventive care, nanoparticles are being incorporated into mouth rinses, toothpaste, and varnishes to reduce plaque formation, enhance remineralization, and combat halitosis.^[4] In periodontology, the nanocoating implant surfaces improve osseointegration and reduces the risks of peri-implantitis by reducing bacterial colonization. Similarly, nanoparticles drug delivery-based systems allow for local and sustained release of medicated agents in periodontal pockets or around infected tissues.^[5]

Selenium nanoparticles (SeNPs) are gaining popularity in medicine due to their novel combination of biological activity, biocompatibility, and lesser toxicity in comparison to other forms of selenium.^[6] SeNPs are immensely popular for their chemical and biological characteristics. Thyroid functioning and antioxidant defense require the trace element selenium. Selenium's bioavailability and reactivity in nanoparticles help combat cancer and other malignancies by encouraging cellular apoptosis. SeNPs are anti-inflammatory, antibacterial, and antioxidant, expanding their medical usefulness.^[6] Selenium oxide nanoparticles (SeO₂ NPs) have many potential applications. They are used in cancer and bacterial infection medicine-based delivery systems due to their oxidative qualities.^[7,8]

The present study aimed to synthesize SeO₂ NPs using a green, eco-friendly technique using *Pleurotus ostreatus* (oyster mushroom) extract as a naturally available reducing and stabilizing agent. Study further sought to characterize the synthesized nanoparticles using physicochemical method and evaluate its antimicrobial efficacy against clinically relevant oral pathogens and their potential biomedical applications.

MATERIALS AND METHODS

This was a laboratory-based, controlled *in vitro* study which aimed to form SeO₂ NPs using an eco-friendly technique employing *P. ostreatus* (oyster mushroom) extract and evaluate its antimicrobial properties. This study was conducted over a 3-month period in the Department of Conservative Dentistry and Endodontics at a university-affiliated dental research laboratory. Ethical clearance was gained from the Institutional Human Ethics Committee (Approval No. SRB/SDC/ENDO-2307/24/465), in conformity with the ethical standards of the Helsinki Declaration and all its amendments.

SYNTHESIS OF SeO₂ NPS

Green synthesis method

SeO₂ NPs were fabricated using an environmentally friendly method which relied on the reducing and stabilizing

properties of *P. ostreatus* (oyster mushroom) extract. This approach allowed the nanoparticles to be produced under gentle conditions without the need for harmful chemicals, making the process both safer and more sustainable.

Materials

Sodium selenite (Na₂SeO₃) was used as the selenium source. An aqueous extract from the plant was prepared and used as the reducing agent. *P. ostreatus* was chosen based on its known antioxidant properties, which are crucial for the reduction of selenium ions to form SeO₂ NPs. Used throughout the experiment for preparing solutions and plant extracts. All glassware used was thoroughly cleaned and dried before use to prevent contamination.

Procedure

The *P. ostreatus* plant leaves were removed, cleaned with distilled water, and air-dried. For 30 min, 20 g of powder was boiled in 100 mL of distilled water. After cooling, the mixture was filtered through Whatman No. 1 filter paper for a clear extract. The extract was stored at 4°C until use. To make a 0.01 M solution, Na₂SeO₃ was dissolved in distilled water. The sodium selenite solution was added drop by drop at room temperature while stirring to incorporate the plant extract. SeO₂ NPs caused the fluid to turn reddish-brown from pale yellow. To reduce selenium ions completely, the reaction mixture was stirred for four hours. Nanoparticles were separated from the solvent by 10,000 rpm centrifugation for 15 min. The nanoparticles were cleaned many times with ethanol and distilled water to remove impurities and unreacted compounds. SeO₂ NPs were dried in a 60°C oven and sealed before characterization.

CHARACTERIZATION TECHNIQUES

To evaluate the physicochemical properties of the synthesized SeO₂ NPs, several advanced characterization techniques were employed. These methods provided insights into the structural, morphological, and chemical properties of the nanoparticles.

X-ray diffraction (XRD)

The phase composition and crystalline structure of the SeO₂ NPs were ascertained using XRD. The desiccated sample was analyzed by the diffraction patterns that were obtained using software.

Fourier transform infrared spectroscopy (FTIR)

FTIR located the SeO₂ NPs' surface functional groups, which may have derived from the plant extract used for synthesis.

Adding potassium bromide to SeO₂ NPs created a pellet. FTIR spectra were taken using a specific model of spectrometer in the 4000–400 cm⁻¹ band. The nanoparticles' spectra showed distinctive absorption bands for functional groups including O-H, C=O, and Se-O, revealing their chemical makeup.

Scanning electron microscopy (SEM)

SEM was used to analyze SeO₂ NPs surface shape and size distribution. Double-sided carbon tape attached dried nanoparticles to an aluminum stub. A sputter coater covered the sample with a thin gold layer to prevent charging during imaging. An SEM model was used to capture SEM images at a certain accelerating voltage, such as 10 kV. Images were analyzed to determine the nanoparticles shape, size, and distribution.

Antimicrobial analysis

Multiple approaches, such as XRD, FTIR, and SEM, are used to characterize synthesized SeO₂ NPs. These approaches reveal the nanoparticles shape, size, size distribution, functional groups, and crystalline structure.

XRD analysis

XRD was used to determine SeO₂ NPs' crystalline structure and phase purity. Figure 5 shows selenium oxide crystallographic plane peaks in the XRD pattern. The XRD pattern presents peaks at specific 2θ values, corresponding to the (101), (110), (200), (211), and (220) planes of crystalline selenium oxide. These peaks match standard diffraction data (JCPDS Card No.), confirming crystalline selenium oxide. SeO₂ NPs crystallite size was determined using the Debye-Scherrer equation:

$$D = \frac{K\lambda}{\beta \cos \theta}$$

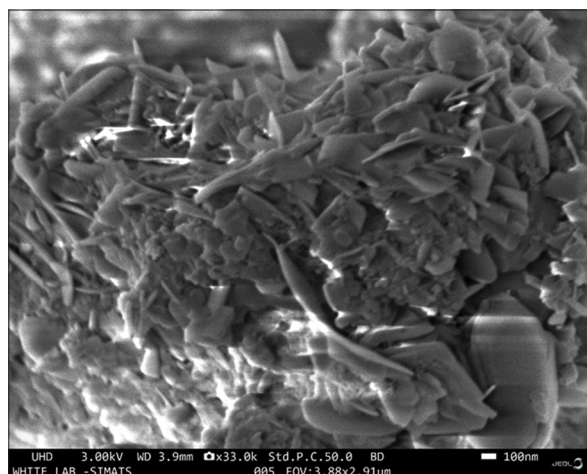


Figure 1: Scanning electron microscopy of samples showing 5,500× magnification

DDD represents crystallite size, KKK is the form factor (usually 0.9), λ measures X-ray wavelength (1.5406 Å for Cu-Kα), β measures peak FWHM, and θ measures Bragg angle. Average crystallite size was [insert size] nm, indicating nanoscale particles. The XRD pattern shows no extra peaks, indicating that the synthesized nanoparticles are phase pure.

FTIR analysis

FTIR spectroscopy was used to identify functional groups on SeO₂ NPs, which may have derived from the plant extract used in their manufacture. Figure 4 shows SeO₂ NPs Fourier transform infrared spectra. The spectrum shows different absorption bands for functional groups. The presence of hydroxyl groups is indicated by a broad band about cm⁻¹, attributed to O-H stretching vibration. Because it stabilized and decreased synthesis, the plant extract likely provided these groups. Selenium oxide is created by stretching vibration of selenium ions, as indicated by a unique peak at wavenumber cm⁻¹. The presence of organic molecules from the plant extract on the nanoparticles' surface is indicated by additional bands at cm⁻¹ and wavenumber cm⁻¹, likely due to C=O and C-H stretching vibrations. Successful surface functionalization of SeO₂ NPs has created functional groups that improve stability and biocompatibility, making them more appropriate for numerous applications.

SEM analysis

SEM was used to analyze selenium oxide nanoparticle surface shape and size distribution. SEM scans, displayed in Figure 3, show that SeO₂ NPs are homogeneous and spherical with an average particle size of up to 10 nm. Well-dispersed particles show less aggregation.

The size distribution histogram from SEM image analysis shows that most nanoparticles are nm. This tight size

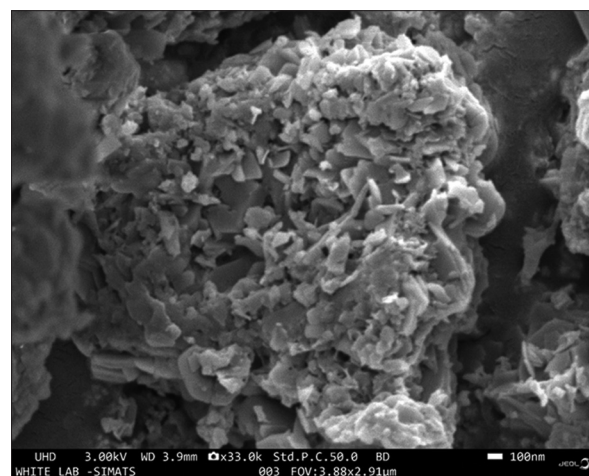


Figure 2: Scanning electron microscopy of samples showing nanoparticles of conical and circular shape at 5,500× magnification

distribution proves that green synthesis produces consistent nanoparticle sizes. The high-resolution SEM pictures show smooth nanoparticle surfaces without flaws or abnormalities. Catalysis and medication transport benefit from this smooth surface.

RESULTS

Multiple characterization methods confirm SeO₂ NPs' eco-friendly production. XRD shows the nanoparticles' nanometer-scale crystalline structure and size. Surface functional groups derived from the plant may have contributed to nanoparticles' stability and biological activity, according to FTIR studies.^[9,10]

SEM investigations precisely depict the nanoparticles' size, shape, and crystallinity, which match SeO₂ NPs features. The nanoparticles' round form and consistent size show that green synthesis can replace chemical synthesis.

The characterization results show that this research's SeO₂ NPs have chemical and structural properties that could be useful in biomedicine, catalysis, and environmental remediation. Green synthesis methods are used for their environmental and safety benefits; therefore, these nanoparticles' particular uses could be studied.

The diffractogram reveals a predominantly crystalline nature, as indicated by sharp and intense diffraction peaks at 2θ values of 23.403°, 28.844°, 31.425°, 43.739°, 45.395°, 51.744°, 55.865°, 61.314°, and 65.382°. The crystalline fraction is calculated to be 70.3%, while the amorphous content is 29.7%. The major diffraction peak at 28.844° corresponds to the characteristic plane of crystalline selenium oxide, suggesting successful nanoparticle formation and high structural order.

We examined the antibacterial activity of SeO₂ NPs against *S. mutans* and *E. faecalis*. Nanoparticle doses of 50 µg/mL, 100 µg/mL, and PC (as a positive control [PC]) were tested

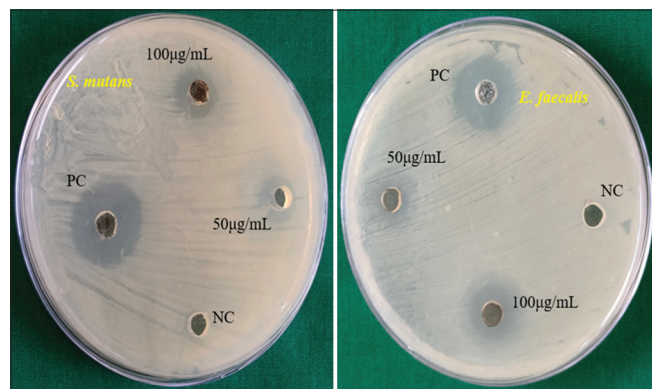


Figure 3: Antibacterial activity assessment for *Streptococcus mutans* (left) and *Enterococcus faecalis* (right) in Mueller-Hinton Agar medium

for efficacy. The inhibition zones were measured at each concentration to determine the nanoparticles' antibacterial activity. Figure 3 data reveal *S. mutans* and *E. faecalis* inhibitory zones (mm) at each concentration level.

In addition, Table 2 includes data with the corresponding standard deviation values.

Antibacterial activity against *S. mutans*

At 50 µg/mL, SeO₂ NPs exhibited modest antibacterial activity against *S. mutans*, with a 7 mm inhibitory zone. Increasing the concentration to 100 µg/mL considerably increased antibacterial activity, expanding the inhibition zone to 11 mm. The PC had the highest antibacterial activity with a 14-mm inhibition zone. Nanoparticles work, but not as well as the PC group's gold standard therapy. The dose-dependent antibacterial effect of SeO₂ NPs against *S. mutans* was found. The inhibition is related to nanoparticle concentration, indicating that higher doses restrict bacterial growth better.

Antibacterial activity against *E. faecalis*

The SeO₂ NPs showed considerable antibacterial effectiveness against *E. faecalis* at 50 µg/mL, with a 10 mm inhibition zone as seen in Table 1. Antibacterial activity increased somewhat at 100 µg/mL, with the inhibition zone expanding to 12 mm. The inhibitory zone at 13 mm for the PC was somewhat greater than that at 100 µg/mL nanoparticles. Antibacterial effectiveness against *E. faecalis* was dose-dependent, like *S. mutans*. Unlike *S. mutans*, nanoparticles reduced bacterial growth, with less difference in inhibition zones at 50 and 100 µg/mL.

Table 1: Zone of inhibition values (in mm) for *S. mutans* and *E. faecalis*

Concentration (µg/mL)	<i>S. mutans</i> (mm)	<i>E. faecalis</i> (mm)
50	7	10
100	11	12
PC	14	13

E. faecalis: *Enterococcus faecalis*, *S. mutans*: *Streptococcus mutans*, PC: Positive control

Table 2: Standard deviation values for *S. mutans* and *E. faecalis*

Concentration (µg/mL)	<i>S. mutans</i> (mm)	<i>S. mutans</i> (SD)	<i>E. faecalis</i> (mm)	<i>E. faecalis</i> (SD)
50	7	1	10	1.1
100	11	1.2	12	1.2
PC	14	1.3	13	0.9

E. faecalis: *Enterococcus faecalis*, *S. mutans*: *Streptococcus mutans*, PC: Positive control

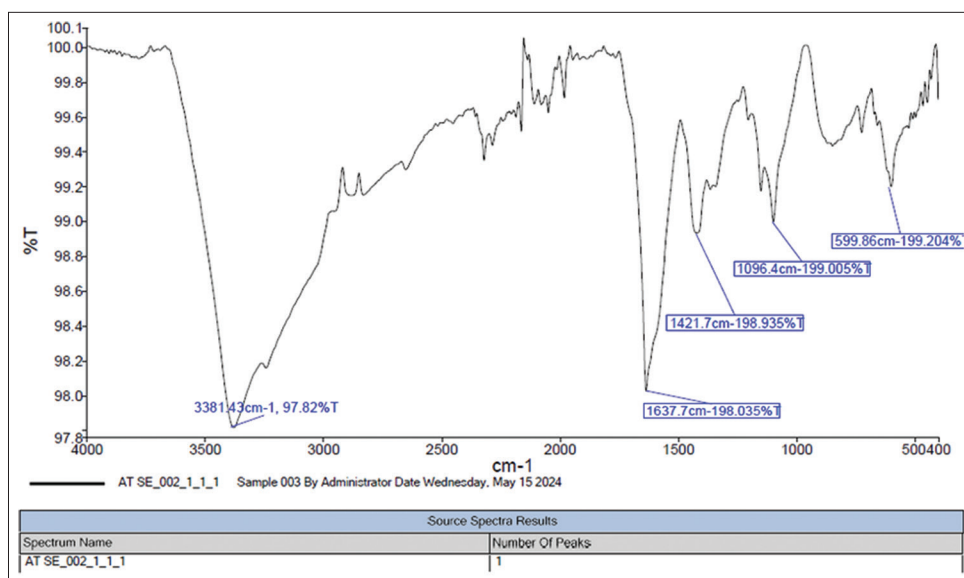


Figure 4: Fourier transform infrared spectroscopy analysis of the nanoparticle

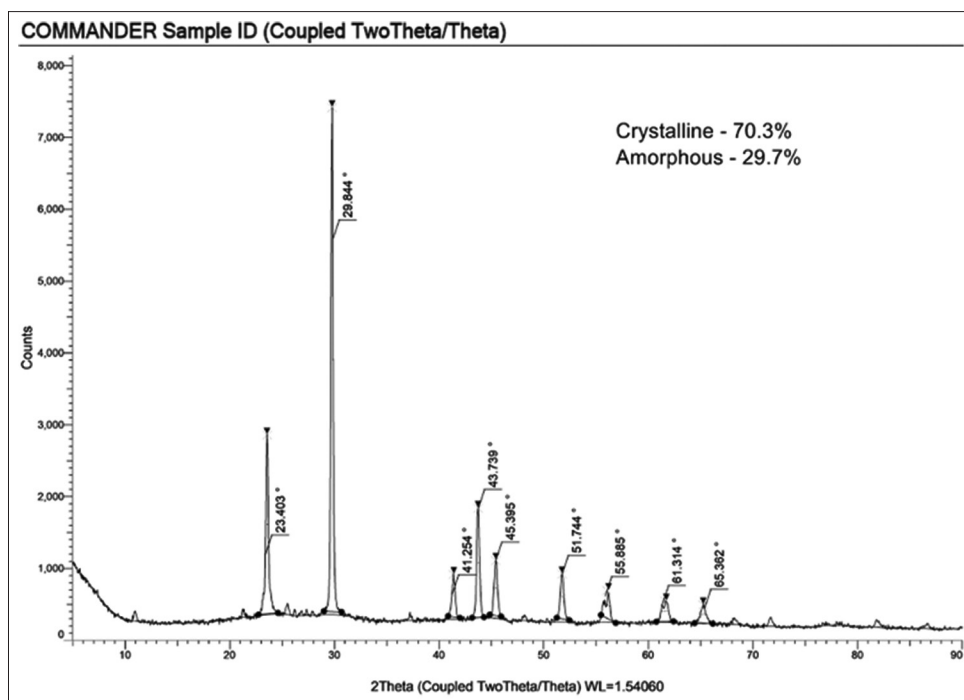


Figure 5: X-ray diffraction pattern of green-synthesized selenium oxide nanoparticles using *P. ostreatus* extract. The diffractogram reveals a predominantly crystalline nature, as indicated by sharp and intense diffraction peaks at 2θ values of 23.403°, 28.844°, 31.425°, 43.739°, 45.395°, 51.744°, 55.865°, 61.314°, and 65.382°. The crystalline fraction is calculated to be 70.3%, while the amorphous content is 29.7%. The major diffraction peak at 28.844° corresponds to the characteristic plane of crystalline selenium oxide, suggesting successful nanoparticle formation and high structural order

Effectiveness of different concentrations as seen in Figure 6

SeO₂ NPs' antibacterial activity increases with concentration. PC, a traditional antibiotic, has the largest inhibitory zones for both bacteria, proving its efficacy. At a dosage of 100 µg/mL, SeO₂ NPs approach the PC's efficiency, suggesting potential as an alternative antibacterial agent. The SeO₂ NPs generated in this study show antibacterial activity against *S. mutans* and *E. faecalis*. The dose-dependent response suggests that

nanoparticles may regulate bacterial growth, especially at higher concentrations. These findings support the concept that SeO₂ NPs could sustainably replace hazardous antibacterial medicines.

DISCUSSION

The green synthesis of SeO₂ NPs using *P. ostreatus* (oyster mushroom) represents a sustainable and biologically active

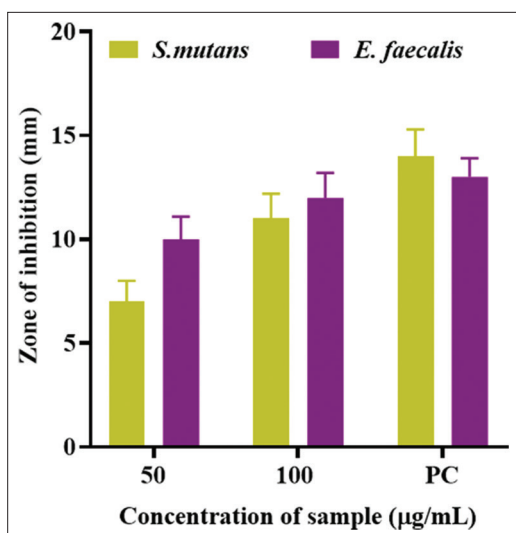


Figure 6: Zone of inhibition in mm of *Streptococcus mutans* and *Enterococcus Faecalis* against the concentration of the synthesized nanoparticle

approach that leverages the reducing and stabilizing potential of mushroom-derived phytochemicals.^[11] This method aligns with the principles of green chemistry, which emphasizes the use of non-toxic, renewable natural resources for nanoparticle fabrication, thereby minimizing environmental impact and improving biocompatibility.^[12]

P. ostreatus is rich in bioactive compounds such as polysaccharides, phenolic, proteins, and flavonoids, which not only act as reducing agents but also serve as effective capping agents. These phytoconstituents enhance the stability and functionality of the synthesized nanoparticles. Similar findings were noted by Cittrarasu *et al.*,^[13] who demonstrated the successful synthesis of SeNPs using *Ceropegia bulbosa* extract, yielding nanoparticles with significant antibacterial, cytotoxic, and photocatalytic activity. Like the plant-based extracts used in their study, the biochemical composition of *P. ostreatus* plays a crucial role in nanoparticle formation and biological activity.^[13]

The SeO₂ nanoparticles synthesized using *P. ostreatus* showed potent antibacterial action, especially against *S. mutans* and *E. faecalis*. This is in accordance with studies which showed that selenium-based nanoparticles synthesized from orange peel waste portrayed strong antibacterial and antibiofilm action against multidrug-resistant strains, showing the potential of waste-derived and cost-effective biomaterials for nanoparticles synthesis.^[14]

The distinct fungal matrix of *P. ostreatus* provides various advantages over plant extracts. Fungal biomolecules offer a more dense and diverse array of enzymes and reducing agents, which contribute to the formation of consistently arranged nanoparticles. Similar biomimetic approaches described by Lashin *et al.*,^[15] who utilized *Ziziphus spina-christi* callus extracts for synthesizing selenium and zinc oxide NPs, showing antimicrobial efficacy.^[15]

The bioactivity of the SeO₂ NPs depends heavily on its physicochemical properties. Factors such as particle morphology, size, and surface chemistry influence their interaction with bacterial membranes. Smaller nanoparticles tend to have a higher surface-to-volume ratio, leading to enhanced ROS formation, membrane disruption, and bacterial cell death.^[16] The mushroom-mediated nanoparticles in this study exhibited these nanoscale properties which are critical and contribute to their antibacterial action.

Furthermore, curcumin-functionalized SeNPs, as explained by Shanmugam *et al.*,^[17] showed varied biological applications, such as in tissue engineering, due to its improved bioavailability and synergism.^[17] This showed the potential for *P. ostreatus*-derived SeNPs to be further functionalized for targeted biomedical therapy.

The immunomodulatory and anti-inflammatory action of SeNPs has been a topic of growing interest. Kurup *et al.*^[18] reviewed the role of metallic nanoparticles in improving the efficacy of anti-inflammatory drugs, revealing that SeNPs might serve dual roles – both as a therapeutic agent and a drug carrier.^[18] This is supported by Vijayaram *et al.*, who revealed diverse applications of green-synthesized metal nanoparticles across bio-medical domains, ranging from antimicrobial coating to cancer therapy.^[19]

In addition, the action of mushrooms for nanoparticle synthesis introduced certain advantages related to scalability, simplicity of biomass cultivation, and lower toxicity.

P. ostreatus is easily cultivable and consumed, making it an economically viable choice which is a safe source for nanoparticle synthesis.^[19,20] These benefits, along with the demonstrated efficacy of SeO₂ NPs, show a promising avenue for future research and development in dental and biomedical-based applications.

Comparisons could also be drawn with SeNPs synthesized from *Withania somnifera*,^[21] *Cleistocalyx operculatus*,^[22] and *Acinetobacter* spp.^[23] All revealed consistent antimicrobial, antioxidant, and anticancer effects. This consistency across diverse biological sources reveals the antibacterial capability of SeNPs when green-synthesized.

The antimicrobial performance of SeO₂ NPs in this study is further supported by Shahmoradi *et al.*,^[24] who combined the SeNPs with photodynamic therapy to improve bacterial biofilm disruption. This combinatorial potential opens future avenues for *P. ostreatus*-mediated SeO₂ NPs into multimodal treatment modality.^[24]

Finally, the biosynthesized nanoparticles showed antioxidant properties, as shown in previous studies like Fan *et al.*,^[25] suggesting a dual role for SeO₂ NPs in both bacterial clearance and cellular protection – making them ideal

candidates for applications in dentistry, wound healing, and tissue regeneration.^[25]

The fabricated SeO₂ NPs show notable antibacterial action against common oral pathogens such as *E. faecalis*, which is a resilient microorganism often seen in endodontic treatment failure. These findings aligned with the work of Hernández-Díaz *et al.*,^[26] who reported that biosynthesized SeNPs showed substantial inhibitory effects against a varied range of clinically relevant bacterial strains, including Gram-positive and Gram-negative organisms. Their study highlighted dose-dependent antibacterial activity and suggested that the bioactivity of SeNPs stems from mechanisms such as oxidative stress induction, membrane disruption, and metabolic interference.^[26]

In the context of oral biofilms, particularly those formed by *E. faecalis*, Shahmoradi *et al.*^[24] demonstrated that SeNPs significantly enhanced the antimicrobial efficacy of photodynamic therapy. Their combined approach disrupted biofilm integrity and increased bacterial susceptibility. The current study, though lacking a combinatorial modality, similarly affirms the standalone efficacy of SeO₂ NPs synthesized through fungal extracts, particularly in disrupting resilient microbial populations. This suggests that *P. ostreatus*-derived nanoparticles may hold potential not only in preventive applications but also as adjuncts in periodontal and endodontic therapy where biofilm resistance is a major challenge.^[24]

The green synthesis strategy employed in this study parallels the work of Vu *et al.*,^[22] who utilized *Cleistocalyx operculatus* leaf extract to synthesize SeNPs. Their work confirmed that biosynthesized SeNPs were biocompatible at low concentrations and exhibited no acute oral toxicity in animal models, thereby supporting the safety of green-synthesized SeNPs for biomedical use. The phytochemical-based synthesis using *P. ostreatus* yielded particles that were stable and effective, and it is likely that the fungal metabolites conferred comparable advantages in terms of less toxicity and improved bioavailability.^[22]

Although selenium and copper are distinct elements, this study by Wu *et al.*^[27] on the green synthesis of copper nanoparticles using *Cissus vitifolia* offered a useful comparison in terms of methodology and observed outcomes. Their copper-based nanoparticles showed potent antioxidant and antimicrobial activity against urinary tract pathogens, and these effects were attributed to both particle morphology along with phytochemical capping agents. The parallel here lies in the biomimetic synthesis route and resultant biological activity. Both copper and SeO₂ NPs synthesized through green methods exhibit enhanced functional properties due to their nano-scale size, surface energy charge, and capping with biologically active molecules.^[27]

The mushroom-based fabrication system used in this study is also advantageous in terms of scalability and environmental

sustainability. Unlike chemically intense synthesis techniques that rely on hazardous reagents, the use of *P. ostreatus* provided a benign, cost-effective alternative that aligned with green chemistry principles. The biological performance of the SeO₂ NPs – characterized by their antibacterial potential – can be largely attributed to the dual role of *P. ostreatus* extracts as both reducing agents and stabilizers.^[28-30]

Taken together, the findings of this study and those of related works support the therapeutic potential of green-synthesized SeNPs. Whether derived from a fungal or plant-based source, SeNPs have shown antibacterial efficacy, led toxicity, and compatibility with biological systems.^[31,32] The added benefit of using an edible mushroom like *P. ostreatus* not only improved the sustainability of the synthesis process but also paved the way for applications in dentistry, such as antimicrobial coatings, irrigants, or adjuncts to regenerative procedures.^[33-36]

CONCLUSION

SeO₂ NPs were synthesized utilizing a green technique and characterized using XRD and antibacterial tests. The major findings reveal that green-chemistry selenium oxide nanoparticles are efficient against *S. mutans* and *E. faecalis*. Nanoparticles were effective at various doses, with the strongest inhibitory zones at higher ones. Green synthesis is necessary to avoid dangerous chemicals and protect the environment. This eco-friendly method promotes biocompatibility and biological applications of nanoparticles. This study shows that sustainable SeO₂ NPs are as effective as conventional ones. SeO₂ NPs' antibacterial properties could be employed in mouthwash or toothpaste to tackle oral disorders like *S. mutans*. Medical equipment coatings or wound dressings may lower infection risk. Future studies may refine the green production process to manage nanoparticle size and form to improve antibacterial efficiency. Nanoparticle synthesis employing plant extracts or other natural resources may reveal the method's versatility and scaling possibilities. To apply these findings, more *in vivo* and molecular action mechanism studies are needed.

AUTHOR'S CONTRIBUTIONS

(1) SA, SDPA, SS: Substantial contribution to the concept or design of the work or the acquisition, analysis, or interpretation of data for the work. (2) SA, SDPA: Drafting the work or reviewing it critically for important intellectual content. (3) SA, SDPA, SS: Final approval of the version to be published. (4) SA, SDPA, SS: Agreement to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

REFERENCES

1. Song W, Ge S. Application of antimicrobial nanoparticles in dentistry. *Molecules* 2019;24:1033.
2. Dilipan E, Sivaperumal P, Kamala K, Ramachandran M, Vivekanandhan P. Green synthesis of silver nanoparticles using seagrass *Cymodocea serrulata* (R. Br.) Asch. and Magnus, characterization, and evaluation of anticancer, antioxidant, and antiglycemic index. *Biotechnol Appl Biochem* 2023;70:1346-56.
3. Saafan A, Zaazou MH, Sallam MK, Mosallam O, El Danaf HA. Assessment of photodynamic therapy and nanoparticles effects on caries models. *Open Access Maced J Med Sci* 2018;6:1289-95.
4. Shanmugam R, Anandan J, Balasubramanian AK, Raja RD, Ranjeet S, Deenadayalan P. Green synthesis of selenium, zinc oxide, and strontium nanoparticles and their antioxidant activity - a comparative *in vitro* study. *Cureus* 2023;15:e50861.
5. Satish S, Sundar S, Kannan S, Ramadoss R, Paneerselvam S, Ramani P. Biosynthesis and characterization of silver nanoparticles derived from ethanol and aqueous extract of tarragon. *J Popul Ther Clin Pharmacol* 2023;30:403-7.
6. Kamala K, Sivaperumal P, Ganapathy D. Avicennia marina mangrove plant mediated selenium nanoparticles and their therapeutic activity against oral pathogens and other biological properties. *J Dent Indones* 2024;31:93-100.
7. Soni J, Revathi D, Dhanraj G, Ramasubburayan R. Bioinspired green synthesis of ZnO nanoparticles by marine-derived *Streptomyces plicatus* and its multifaceted biomedical properties. *Microb Pathog* 2024;193:106758.
8. Majeed A, Javed F, Akhtar S, Saleem U, Anwar F, Ahmad B, *et al.* Green synthesized selenium doped zinc oxide nano-antibiotic: Synthesis, characterization and evaluation of antimicrobial, nanotoxicity and teratogenicity potential. *J Mater Chem B* 2020;8:8444-58.
9. Shaaban M, El-Mahdy AM. Biosynthesis of Ag, Se, and ZnO nanoparticles with antimicrobial activities against resistant pathogens using waste isolate *Streptomyces enissocaesilis*. *IET Nanobiotechnol* 2018;12:741-7.
10. Hatami R, Javadi A, Jafarizadeh-Malmiri H. Effectiveness of six different methods in green synthesis of selenium nanoparticles using propolis extract: Screening and characterization. *Green Process Synth* 2020;9:685-92.
11. Ghaderi RS, Adibian F, Sabouri Z, Davoodi J, Kazemi M, Jamehdar SA, *et al.* Green synthesis of selenium nanoparticle by *Abelmoschus esculentus* extract and assessment of its antibacterial activity. *Mater Technol* 2022;37:1289-97.
12. Raman S, Kasirajan S, Chinnapandi B, Ramalingam S, Vijayakumar R, Arumugam P. Luminescent biogenic selenium nanoparticles from *Indigofera aspalathoides*: Green synthesis, characterization and *in vitro* evaluation of antioxidant and hepatoprotective potential. *Luminescence* 2025;40:e70101.
13. Citrarasu V, Kaliannan D, Dharman K, Maluventhen V, Easwaran M, Liu WC, *et al.* Green synthesis of selenium nanoparticles mediated from *Ceropegia bulbosa* Roxb extract and its cytotoxicity, antimicrobial, mosquitocidal and photocatalytic activities. *Sci Rep* 2021;11:1032.
14. Lashin I, Hasanin M, Hassan SA, Hashem AH. Green biosynthesis of zinc and selenium oxide nanoparticles using callus extract of *Ziziphus spina-christi*: Characterization, antimicrobial, and antioxidant activity. *Biomass Convers Biorefin* 2023;13:10133-46.
15. Salem SS, Badawy MS, Al-Askar AA, Arishi AA, Elkady FM, Hashem AH. Green biosynthesis of selenium nanoparticles using orange peel waste: Characterization, antibacterial and antibiofilm activities against multidrug-resistant bacteria. *Life (Basel)* 2022;12:893.
16. Borowska M, Jankowski K. Green synthesis of selenium nanoparticles: Characterization and application. In: *Handbook of Greener Synthesis of Nanomaterials and Compounds*. Netherlands: Elsevier; 2021. p. 171-190.
17. Veeraraghavan VP, Gayathri R, Shanmugam SB, Sankaran K. Curcumin-coated *Orthosiphon stamineus* leaf extract-based selenium nanoparticles for potential tissue engineering applications. *Texila Int J Public Health*. 2023. doi: 10.21522/TIJPH.2013.SE.23.01.Art005
18. Kurup M, Kumar M, Ramanathan S, Rajappa MC. The biogenetic synthesis of metallic nanoparticles and the role they play in the anti-inflammatory drug treatment. *Curr Drug Discov Technol* 2024;21:e180723218848.
19. Ijayaram S, Razafindralambo H, Sun YZ, Vasantharaj S, Ghafarifarsani H, Hoseinifar SH, *et al.* Applications of green synthesized metal nanoparticles - a review. *Biol Trace Elem Res* 2024;202:360-86.
20. Subha T, Srilatha M, Naveen P, Thirumalaisamy R. Green synthesis, characterization and optimization of silver nanoparticles from *Carica papaya* using Box Behnken design and its activity against dental caries causing *Streptococcus* sp. *Chem Data Collect* 2024;51:101139.
21. Alagesan V, Venugopal S. Green synthesis of selenium nanoparticle using leaves extract of *Withania somnifera* and its biological applications and photocatalytic activities. *Bio Nano Sci* 2019;9:105-16.
22. Vu TT, Nguyen PT, Pham NH, Le TH, Nguyen TH, Do DT, *et al.* Green synthesis of selenium nanoparticles using *Cleistocalyx operculatus* leaf extract and their acute oral toxicity study. *J Compos Sci* 2022;6:307.
23. Wadhwani SA, Gorain M, Banerjee P, Shedbalkar UU, Singh R, Kundu GC, *et al.* Green synthesis of selenium nanoparticles using *Acinetobacter* sp. SW30: Optimization, characterization and its anticancer activity in breast cancer cells. *Int J Nanomedicine* 2017;12:6841-55.
24. Shahmoradi S, Shariati A, Zargar N, Yadegari Z, Asnaashari M, Amini SM, *et al.* Antimicrobial effects of selenium nanoparticles in combination with photodynamic therapy against *Enterococcus faecalis* biofilm. *Photodiagnosis Photodyn Ther* 2021;35:102398.

25. Fan D, Li L, Li Z, Zhang Y, Ma X, Wu L, *et al.* Biosynthesis of selenium nanoparticles and their protective, antioxidative effects in streptozotocin induced diabetic rats. *Sci Technol Adv Mater* 2020;21:505-14.
26. Hernández-Díaz JA, Garza-García JJ, León-Morales JM, Zamudio-Ojeda A, Arratia-Quijada J, Velázquez-Juárez G. Antibacterial activity of biosynthesized selenium nanoparticles using extracts of *Calendula officinalis* against potentially clinical bacterial strains. *Molecules* 2021;26:5929.
27. Wu S, Rajeshkumar S, Madasamy M, Mahendran V. Green synthesis of copper nanoparticles using *Cissus vitifolia* and its antioxidant and antibacterial activity against urinary tract infection pathogens. *Artif Cells Nanomed Biotechnol* 2020;48:1153-8.
28. Gandhimathi KA, Francis AA, Rengasamy G, Vishnu Priya V, Sankaran K. Quercetin-coated biogenic titanium oxide nanoparticles: Synthesis, characterization, and *in-vitro* biological studies. *Part Sci Technol* 2025;43:687-97.
29. Farhana SP, Francis AP, Gayathri R, Sankaran K, Veeraraghavan VP. Synthesis and characterization of zirconium oxide nanoparticles based on *Hemidesmus indicus* extract: Evaluation of biocompatibility and bioactivity for prosthetic implant coatings. *J Adv Oral Res* 2024;15:45-51.
30. Ramamoorthy K, Uma Maheswari TN, Muthupandian S, Pillai DS. One-pot green synthesis of strontium and zirconium nanoparticles: An *in-vitro* study of their antioxidant and cytotoxic activity. *J Clin Diagn Res* 2025;19:FC01-5.
31. Hameed MS, Delphine PS, Shanmugam R, Raghu S. Synthesis and characterization of nanosilica and its interaction with antibiotics: Nanosilica antibiotic-coated variants. *J Int Oral Health* 2024;16:386-93.
32. Tamanna IS, Gayathri R, Sankaran K, Veeraraghavan VP, Francis AP. Eco-friendly synthesis of selenium nanoparticles using *Orthosiphon stamineus* leaf extract and its biocompatibility studies. *BioNano Sci* 2024;14:37-44.
33. Valan AS, Hema M. Eco-friendly synthesis of copper sulphate nanoparticles using *Citrus sinensis* extract and their antimicrobial properties: An *in-vitro* study. *J Clin Diagn Res* 2024;18:ZC42-6.
34. Chandran N, Ramesh S, Adarsh VJ, Rakesh VK. Comparative evaluation of contact angle in biosynthesized and chemically synthesized silver nanoparticle-based root canal irrigants: An *in-vitro* study. *J Clin Diagn Res* 2024;18:ZC58-62.
35. Sankaranarayanan SA, Roy A, Surya M. Green synthesis of *Camellia sinensis*-mediated selenium-doped vitamin E and chitosan nanoparticles along with evaluation of their anti-inflammatory and anticancer activity: An *ex-vivo* study. *J Clin Diagn Res* 2025;19:FC22-6.
36. Kothari V, Rajaraman V, Ariga P, Sekaran S, Ganapathy DM. Antibacterial efficacy and antioxidant potential of hafnium-coated titanium implants: An *in-vitro* assessment. *J Clin Diagn Res* 2025;19:ZC21-5.

Source of Support: Nil. **Conflicts of Interest:** None declared.